

A monocenter retrospective study of serological, histological and genetic characteristics of celiac disease in Northern Central Italy

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ABSTRACT

Objective: To examine the seroprevalence, correlates and characteristics of Celiac disease (CD) in a population sample of a Northern Central Area of Italy, by a monocenter retrospective study. **Methods:** Between 2006 and 2010, serum samples of 9371 subjects (age range 6 months to 91 years) were screened for tissue transglutaminase IgA antibodies (IgA-tTG) by the Immunologia-Allergologia Unit of AUSL1 Massa-Carrara, an area with a population of approximately 150,000. Endomysial IgA antibodies (EMA), HLA typing and small-bowel biopsy were also performed when indicated. **Results:** Of the 9371 subjects, 269 (2.87%) had positive antibody tests. The population was divided into several age groups and the highest prevalence (5.63%) was found in the 6 months - 3 years group. The prevalence of IgA-tTG positivity was double in females compared to males. All IgA-tTG positive patients that were genotyped carried HLA-DQ2 or DQ8, none was $\alpha 5$ positive only. In positive IgA-tTG sera, levels of IgA-tTG were significantly higher in EMA positive than in EMA negative sera ($p < 0.001$) both in children and in adults. Ninety-five/269 IgA-tTG positive subjects underwent biopsy. IgA-tTG levels were different according to the histological degree of the lesion. When EMA were evaluated in IgA-tTG positive subjects the number of EMA negative sera was significantly higher in adults than in children. **Conclusions:** In summary, this study provides a monocenter retrospective analysis of serological, histological and genetic parameters of subjects with suspicion of CD in an area of Northern Central Italy from 2006 to 2010.

Keywords: Celiac Disease; Anti-Endomysium Antibodies; Anti-Transglutaminase Antibodies; Epidemiology; HLA-DR

1. INTRODUCTION

Celiac disease (CD) is an autoimmune enteropathy characterized by chronic intestinal inflammation resulting in villous atrophy and flattening of the small intestinal mucosa. CD develops in genetically predisposed individuals in response to the dietary ingestion of wheat gluten and similar proteins in barley and rye. Originally considered a rare malabsorption syndrome in childhood, CD is now recognized as a common disorder that may arise at any age, with growing proportion of new cases diagnosed in adults and patients with extraintestinal manifestations, such as esophageal reflux, osteoporosis, hypertransaminasemia and neurological symptoms [1,2]. Although CD is one of the most common lifelong diseases in western countries, most affected individuals remain undiagnosed [3]. This is apparently because many patients have atypical symptoms or none at all. The disease is characterized by the production of anti-tissue transglutaminase (IgA-tTG) and anti-endomysial IgA (EMA) antibodies [4-6]. Serological screening for the presence of these autoantibodies in individuals with characteristic symptoms of CD or with associated conditions is usually the initial step in detecting new cases. Although IgA-tTG and EMA appear to be good markers of the active phase of the disease, the definitive diagnosis requires a small-bowel biopsy showing the typical histological abnormalities (villous atrophy, crypt hyperplasia and intraepithelial lymphocytes) [7]. Population based studies have shown that the prevalences of CD in Europe and North America

range from 0.7% to 2% [3,8-10]. The aim of this mono-center retrospective study was to investigate the serological, histological and genetic characteristics of subjects with CD suspicion who attempt to the Massa Hospital, Northern Central Italy, from 2006 up to 2010.

2. METHODS

2.1. Study Population

Subjects who underwent serological investigations by the Immunologia Allergologia Unit of Azienda USL1 Massa Carrara, between January 2006 and December 2010, because of suspicion of CD, were included in the present study. We examined 14247 subjects residing in the Massa Carrara area, an area with a population of approximately 150,000. Multiple accesses for the same subjects have been observed, therefore in the present study it has been decided to include data regarding only one access according to the following criteria: results of the last access were used for subjects with more accesses all negative for IgA-tTG; results of the first access positive for IgA-tTG were used for subjects with more accesses one of which positive for IgA-tTG. On the basis of this recruitment criterion the data analysis was restricted to 9371 out of 14247 initial ones, of which 3234 male and 6137 female.

2.2. Autoantibodies Measurements

IgA-tTG were measured by EliA Celikey (Phadia, Friburg, Germany) using cutoffs as recommended by the manufacturer [11]. Results were expressed as arbitrary units (EliA U/ml) derived from standard curves of serial dilutions of calibrators.

Presence of EMA was measured by an immunofluorescence method using monkey esophagus as substrate (Euroimmun, Germany) [12]; a positive fluorescence at serum dilutions equal to or greater than 1:10 was considered positive. The EMA assay was performed when required by physician.

2.3. Intestinal Histology

Histological evaluations were performed blindly by a single skilled operator. The biopsy's grading was made according to the Marsh's classification modified by Oberhuber *et al.* [13].

2.4. HLA Haplotypes and Nomenclature

Genomic DNA was extracted from whole blood samples using Sample Prep Trombo (Diatech, Italy). The HLA typing was performed by a polymerase chain reaction (PCR) with a commercially available kit (Protrans HLA Celiac Disease Domino System, Protrans, Ketsch, Germany). The HLA kit contained multiplex PCRs for two

HLA-class II markers, the HLA-DQ2 (DQA1*0501 and DQB1*0201) and the HLA-DQ8 (DQA1*0301 and DQB1*0302) and for beta globin protein as internal control. The obtained amplicons were resolved on 2% agarose gel and stained using ethidium bromide.

The term DQ2 belongs to the serologic HLA nomenclature, and it specifies an epitope on the $\beta 2$ chain. However with time, in CD, the term has usually referred to a particular $\alpha 5\beta 2$ DQ2 dimer encoded by DQA1*05 and DQB1*02 alleles. Therefore, DQ2 is used to indicate subjects carrying both the alleles, whereas individuals DQA1*05 negative/DQB1*02 positive, in which the $\beta 2$ chain forms dimer with a different α chain, are simply named $\beta 2$. A single or double dose of DQB1*02 is indicated as B1*02/X or B1*02/02, respectively. The phenotype coded by DQA1*05 allele in absence of DQB1*02 is designed $\alpha 5$.

2.5. Statistical Analysis

Discrete variables were compared, by group, using chi square test. When comparing the serological assays by age groups, the Mann-Whitney nonparametric U test was used and results were presented as median concentrations. Values of $p < 0.05$ were considered to be significant.

3. RESULTS

The general demographic characteristics of the screened subjects in the years, the age distribution and the number of new IgA-tTG identified patients are summarized in **Table 1**.

Of the 9371 screened subjects, 269 (2.87%) were IgA-tTG positive. When the population was subdivided into age groups, the highest prevalence (5.63%) was found in the 6 months - 3 years group. Moreover, subdividing the population into children (6 months - 15 years) and adults (>15 years) it was observed that 98 out of 2625 (3.73%) children and 171 out of 6746 (2.53%) adults were IgA-tTG positive, respectively. This significant difference ($p < 0.001$) indicates that the IgA-tTG seropositivity is found mostly in pediatric age.

The gender distribution of the studied population is reported in **Table 2**. The pediatric population is equally divided into female and male subjects; on the contrary, in the adult group, female subjects are more than double compared to male subjects. The IgA-tTG prevalence was therefore calculated considering the pediatric population: 66 out of 1317 (5.01%) female subjects and 32 out 1308 (2.44%) males were IgA-tTG positive respectively, with a ratio female/male = 2/1.

Of the 9371 subjects, 476 were genotyped for HLA-DRB1, DQA1 and DQB1 loci. As it has not always been possible to have certain information regarding the clinical reason of the genotype request *i.e.* first degree of re-

Table 1. IgA-tTG positivity.

Year	N° of screened subjects	Incidence of IgA-tTG positive subjects	
		N°	%
2006	2290	68	2.96
2007	2009	45	2.23
2008	1477	42	2.84
2009	1840	60	3.26
2010	1755	54	3.07
Prevalence of IgA-tTG positive subjects			
Age group	N° of screened subjects	N°	%
6 - 36 months	834	47	5.63
4 - 10 years	1094	32	2.92
11 - 15 years	697	19	2.72
16 - 30 years	1681	56	3.33
31 - 45 years	2227	71	3.18
46 - 60 years	1473	28	1.90
61 - 80 years	1174	14	1.19
>80 years	191	2	1.04

Table 2. IgA-tTG prevalence per gender.

	Gender	N° subjects	IgA-tTG positive subjects	
			N°	%
6 months - 15 years	Female	1317	66	5.01
	Male	1308	32	2.44
	Total	2625	98	3.73
>15 years	Female	4820	121	2.51
	Male	1926	50	2.59
	Total	6746	171	2.53

relationship, or clinical picture suggestive of CD, we evaluated only the haplotype frequency of the studied population (**Table 3**).

Of all subjects positive for HLA CD predisposing alleles, 57% carried DQ2 and/or DQ8 heterodimers, 16% had $\beta 2$ chain and 27% was $\alpha 5$ positive. All IgA-tTG patients that were genotyped carried HLA-DQ2 or DQ8, none was $\alpha 5$ positive only.

Out of 9371 subjects, 269 were positive for IgA-tTG and levels of IgA-tTG were significantly higher in children than in adults (median value 89 U/ml and 38 U/ml respectively, $p < 0.001$, data not reported). Results of IgA-tTG sera screened for EMA are reported in **Table 4**; levels of IgA-tTG were significantly higher in EMA positive than in EMA negative sera ($p < 0.001$) both in children and in adults.

Ninety-five subjects out of 269 IgA-tTG positive sub-

Table 3. HLA haplotypes.

	General population	IgA-tTG and EMA positive subjects
DR3-DQ2/-	120	7
DR3-DQ2/DR7-DQ2	30	3
DR3-DQ2/DR5-DQ7	2	0
DR5-DQ7/DR7-DQ2	35	3
DR3-DQ2/DR3-DQ2	1	3
DR4-DQ8	45	2
DQA1*05	111	0
DQB1*02/02 and DQB1*02/X	64	5
HLA other than DQ2/DQ8	68	0

Table 4. IgA-tTG levels in EMA screened sera.

	EMA positive		EMA negative	
	N° subjects	IgA-tTG median (U/ml)	N° subjects	IgA-tTG median (U/ml)
6 months - 15 years	59	99	23	21
>15 years	60	61	57	28

jects accepted to undergo biopsy.

Levels of IgA-tTG varied according to the histological degree of the lesion, as reported in **Table 5**.

IgA-tTG levels correlate well with the severity of the lesion as already reported [14,15].

As reported in Methods, EMA was performed only when expressly required, therefore 199 out of 269 IgA-tTG positive sera were investigated for EMA (**Table 6**).

It was found that the number of EMA negative sera was significantly higher in adults than in children both in the studied population and in the 77 samples in which biopsy was performed.

4. DISCUSSION

The prevalence of CD in the studied population of this area of Northern Central Italy is 2.87%. This result is not in contrast with that of 0.7% - 2% reported in previous studies on general population [3,8-10] as we investigated a population of CD suspected subjects. We estimated the prevalence of CD assuming that all IgA-tTG positive subjects had CD. The lack of confirmation of the diagnosis by means of an intestinal biopsy in all IgA-tTG positive subjects may represent a potential limitation of the study.

However, this assumption is supported by previous studies [16-18] and by our observation that the 87% of IgA-tTG positive subjects who underwent intestinal biopsy had a lesion consistent with CD; the remaining 13% of subjects had type I-II lesions in the presence of gas-

Table 5. IgA-tTG level in biopsied subjects.

Mucosal histopathology Marsh Oberhuber classification	IgA-tTG level	
	N° subjects	Median (U/ml)
Type 0	0	
Type I	6	18
Type II	6	
Type IIIa	19	54
Type IIIb	23	67
Type IIIc	41	128

Table 6. EMA in positive IgA-tTG subjects.

Mucosal histopathology Marsh Oberhuber classification	All subjects			
	Adults		Children	
	EMA negative	EMA positive	EMA negative	EMA positive
	32	85	7	75
Mucosal histopathology Marsh Oberhuber classification	Biopsied subject			
	Adults		Children	
	EMA negative	EMA positive	EMA negative	EMA positive
Type 1	3	6		1
Type 2	1	1		1
Type IIIa	2	8		
Type IIIb	2	15	1	1
Type IIIc	4	19	1	11

trointestinal symptoms and HLA haplotypes compatible with CD. The prevalence of CD is almost constant during the time-span examined, being 2.96% in 2006 and 3.07% in 2010.

In this study, a bimodal pattern of CD diagnosis was observed with a first peak at 6 to 36 months and a second one at 16 to 45 years.

The total prevalence of CD decreases in a statistically significant manner in the pediatric population from the highest value of 5.63% to 2.72% of the 6 - 36 months and 11 - 15 years respectively, and the decreasing trend could also be seen in the adult population with the highest value of 3.33% and the lowest one of 1.04% in the 16 - 30 and >80 age groups respectively.

Overall, the prevalence of CD is twice as frequent among females as compared to males, possibly because the necessary HLA haplotypes, DQ2/DQ8, are more frequent in female than in male patients [19]. Inside the adult population the request of IgA-tTG is much more frequent for females especially during the third to fourth decades of their life; this could be explained either because females tend to seek medical care more often than males [10] or because pregnancy or other hormonal disorders could trigger CD symptoms [20]. Several studies

indicate that CD can develop also in the elderly [21-23] with variable symptoms ranging from silent disease, to vague abdominal complaints, to anemia. These limited symptoms could also lead to a delay in diagnosis. In the studied population, 15% of CD diagnostic requests involved subjects over 60 years of age, thus indicating that in our study area the level of attention to this disease by the physicians is very high throughout the patient's life.

According to our HLA genotype results, in the studied population the mostly represented genotypes related to CD are DQ2 and β 2 (62%) with DQ8 at 11%; inside IgA-tTG patients DQ2 and β 2 genotypes arise to 91% while DQ8 doesn't change significantly (9%) and none was α 5 positive only or HLA different from DQ2/DQ8, thus confirming what reported in previous studied [24].

Serological analysis for IgA-tTG shows in the 269 positive subjects that the level of antibodies was significantly higher in the pediatric population than in adults and it shows to correlate well with severity of biopsy as already reported [25]. Moreover when IgA-tTG positive sera were tested for EMA it has been found that level of IgA-tTG significantly correlates with presence of EMA either in children than in adults.

The prevalence figure (18%) for EMA negative CD in the biopsied population was slightly higher when compared with that of 15% reported in other studies [26,27].

Almost all EMA negative patients were adults (12 versus 2). Further studies would be required to explain this finding, however during a longstanding immune reaction, antibodies with increasing avidity are produced which makes it understandable why older patients with CD may have lower IgA-tTG and EMA serum levels than the younger ones; thus longstanding CD might even result in seronegativity.

In summary, this paper provides for the first time a picture of CD in an area of Northern Central Italy, one of the countries with the highest incidence of CD in the world [28].

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