

Pyroglutamated Apelin-13 Inhibits Lipopolysaccharide-Induced Production of Pro-Inflammatory Cytokines in Murine Macrophage J774.1 Cells

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Abstract

Apelin, recently identified as an endogenous ligand of the orphan G protein-coupled receptor APJ, has multiple pathophysiological properties. In the present study, we investigated whether pyroglutamated apelin-13 ([Pyr1]-apelin-13), the most highly active isoform among the mature apelin peptide family, modulates the effect of bacterial lipopolysaccharide (LPS) on cytokine induction in a murine macrophage-like cell line, J774.1 cells. J774.1 cells expressed the APJ protein in a stationary state, and the expression of APJ was not affected by LPS stimulation. No significant effect of [Pyr1]-apelin-13 treatment alone was observed on the proliferation or cytokine production of J774.1 cells in the stationary state. However, prior to LPS stimulation, pretreatment with [Pyr1]-apelin-13 for 16 h significantly diminished mRNA expression and protein secretion of inflammatory cytokine interleukin-6, which was confirmed by RT-PCR and ELISA, respectively. Western blot analysis revealed that the phosphorylation of p38 mitogen-activated protein kinase and c-Jun N-terminal kinase, but not extracellular signal-regulated kinase, which was induced by LPS, significantly decreased in [Pyr1]-apelin-13-pretreated J774.1 cells compared with untreated cells. These observations suggest that [Pyr1]-apelin-13 functions as a negative regulator of LPS-mediated pro-inflammatory responses in macrophages.

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Keywords

Apelin, Macrophage, Inflammation

1. Introduction

APJ, which is highly conserved in vertebrates, is similar to the angiotensin II type 1 receptor, but does not bind angiotensin II. Apelin, originally identified in bovine stomach extracts, is an endogenous ligand of the orphan Gi-G protein-coupled receptor APJ. Apelin has several mature forms. Apelin is initially synthesised as a 77-amino acid pre-propeptide that is cleaved to yield bioactive peptides of 36, 17, 13 and 12 amino acids in length [1]. These mature forms of apelin lack cysteine residues, suggesting that they are monomeric [2]. Among the active forms of the apelin family, the pyroglutamated isoform of apelin ([Pyr1]-apelin-13) has the highest abundance and activity based on its resistance to degradation [1]. The sequence of the C-terminal apelin-13-peptide is conserved in most vertebrates, suggestive of evolutionary conservation and having an important function [3].

APJ and apelin are found in a wide variety of tissues, including brain, heart, lung, kidney, gastrointestinal tract, retina and mammary glands [4]-[7]. Growing evidence suggests that apelin has many physiological functions, such as fluid homeostasis, food intake, cell proliferation, blood pressure regulation, cardiovascular function, angiogenesis and glucose utilization. Thus, the apelinergic system plays an important role in central and peripheral homeostatic regulation [8]-[11].

Apelin is mainly produced and secreted by adipocytes and stimulates glucose uptake through the PI3K/AKT pathway [12]. In clinical studies, serum apelin levels were positively correlated with body mass index, and the levels increased in patients with impaired glucose intolerance or type 2 diabetes [13] [14]. Immunologically, apelin has been shown to suppress cytokine production by mouse spleen cells in response to T-cell receptor/CD3 cross-linking [15]. Apelin inhibits cholinergic activity by decreasing acetylcholine synthesis and release through the downregulation of choline acetyltransferase mRNA expression in the leukemic T-cell line, MOLT-3 [16]. Although immunological studies on isoforms of the apelin peptide have been performed, the role of apelin in host defense against bacterial pathogens remains unclear.

In the present study, we investigated the effect of [Pyr1]-apelin-13 on immunological and inflammatory reactions in lipopolysaccharide (LPS)-stimulated J774.1 murine macrophage-like cells.

2. Methods

2.1. Reagents and Antibodies

Synthetic pyroglutaminated apelin-13 ([Pyr-1]-apelin-13) was purchased from the Peptide Institute, Inc. (Osaka, Japan). LPS derived from *Escherichia coli* (serotype O111:B4) and anti- β -actin monoclonal antibody were obtained from Sigma-Aldrich (St. Louis, MO, USA). Anti-mouse APJ antibody was acquired from GeneTex, Inc. (Irvine, CA, USA). Anti-mouse phosphorylated p38 mitogen-activated protein kinase (p-p38 MAPK), phosphorylated p44/42 MAPK (p-Erk1/2) and phosphorylated stress-activated protein kinase/Jun-amino-terminal kinase (p-SAPK/JNK) antibodies were purchased from Cell Signaling Technology, Inc. (Boston, MA, USA). 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) was purchased from Molecular Probes (Carlsbad, CA, USA).

2.2. Cell Culture

The murine macrophage cell line J774.1 was obtained from the Japanese Cancer Research Resources Bank (Tokyo, Japan) and murine macrophage cell line RAW264.7 was obtained from the American Type Culture Collection (Manassas, VA, USA). J774.1 and RAW264.7 were maintained at 37°C in RPMI-1640 medium (Gibco-BRL, Grand Island, NY, USA) supplemented with 10% heat-inactivated fetal bovine serum (FBS), penicillin G (100 U·mL⁻¹) and streptomycin (100 μ g·mL⁻¹) in an atmosphere of 5% CO₂ in air. Cells (2×10^5 cells·mL⁻¹) were then cultured with RPMI-1640 medium containing 5% FBS in a 6-well plastic plate and preincubated with [Pyr-1]-apelin-13 for 16 h. Thereafter, cells were cultured with or without LPS (10 ng·mL⁻¹), after which samples were harvested for assays.

2.3. Quantitative Real-Time Reverse Transcription (RT)-PCR

Total RNA was isolated from cells using the RNeasy[®] Mini Kit (Qiagen, Valencia, CA, USA). Samples were reverse transcribed with q-Script cDNA Supermix reagents (Quanta BioSciences, Gaithersburg, MD, USA) and amplified for 30 min at 42°C. For real-time RT-PCR, the PCR products were detected using FAST SYBR[®] Green Master Mix (Applied Biosystems, Foster City, CA, USA) using the following primer sequences: *GAPDH* forward, 5'-GACGGCCGCATCTTCTTGA-3' and reverse, 5'-CACACCGACCTTACCATT-3'; *IL-1 β* forward, 5'-AGTTGACGGACCCCAAAAGA-3' and reverse, 5'-GGACAGCCCAGGTCAAAGG-3'; and *IL-6* forward, 5'-CCACGGCCTTCCCTACTTC-3' and reverse, 5'-TTGGGAGTGGTATCCTCTGTGA-3'. Thermal cycling and fluorescence detection were performed using a StepOne[™] Real Time PCR System (Applied Biosystems). The mRNA expression levels were quantified according to the $\Delta\Delta$ CT method, normalised against *GAPDH* expression and presented relative to control levels.

2.4. Western Blot Analysis

Total protein was extracted using Cell Lysis Buffer (Cell Signaling Technology, Inc.) containing a protease inhibitor cocktail (Thermo Fisher Scientific, Waltham, MA, USA). Protein concentrations were determined using the DC[™] Protein Assay Kit (Bio-Rad, Hercules, CA, USA). The same amounts of total protein were loaded in each lane for sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) using 12.5% gels, and were then transferred to polyvinylidene difluoride membranes (Millipore Corp., Bedford, MA, USA). After blocking for 30 min in Blocking One (Nacalai Tesque, Inc., Tokyo, Japan), the membranes were incubated overnight with diluted primary antibodies at 4°C. Antibody binding was detected using horseradish peroxidase-conjugated secondary antibodies. Chemiluminescence was produced using ECL Prime Western Blotting Reagent (GE Healthcare, Little Chalfont, UK) and detected digitally with GelDoc XR Plus (Bio-Rad).

2.5. Determination of IL-6 Concentrations

A sandwich ELISA was performed using the mouse IL-6 ELISA kit (Thermo Fisher Scientific) according to the manufacturer's protocol. Conditioned media were diluted between 1:10 and 1:50.

2.6. Statistical Analysis

Statistical analysis was conducted with JMP software, version 9.0.2. Data are presented as means \pm SD. Groups were compared using the two-tailed Students *t*-test for parametric data. A value of $P < 0.05$ was considered statistically significant.

3. Results

3.1. Expression of the Apelin Receptor APJ in Macrophage Cell Lines

To investigate the immune function of apelin in macrophages, we first confirmed the expression of APJ in well-known macrophage cell lines, J774.1 and RAW 264.7 cells. Apelin administration had no effect on cell proliferation based on MTT assays (data not shown). Western blotting analysis revealed that both cell lines expressed APJ protein in their stationary state (**Figure 1**). Treatment with [Pyr1]-apelin-13 for 16 h had no effect on APJ expression. In addition, LPS administration did not affect the amount of APJ expression in the two cell lines.

3.2. Effect of Apelin on LPS-Induced mRNA and Protein Expression of Pro-Inflammatory Cytokines

We next examined the effect of [Pyr1]-apelin-13 on the LPS-stimulated inflammatory response in J774.1 cells. J774.1 cells were treated with 1 μ M [Pyr1]-apelin-13 for 16 h and were then administered 10 ng·mL⁻¹ LPS for an additional 2 - 4 h. Real-time RT-PCR analysis revealed that pretreatment with [Pyr1]-apelin-13 decreased LPS-induced *IL-1 β* and *IL-6* mRNA expression (**Figure 2(a)** and **Figure 2(b)**, respectively). The LPS-induced expression of *IL-1 β* and *IL-6* mRNA were suppressed by 65.2% \pm 4.2% and 63.3% \pm 2.6%, respectively, by [Pyr1]-apelin-13 pretreatment in J774.1 cells. In addition, ELISA analysis revealed that pretreatment with [Pyr1]-apelin-13 significantly suppressed LPS-induced *IL-6* secretion from J774.1 cells (**Figure 3**). These re-

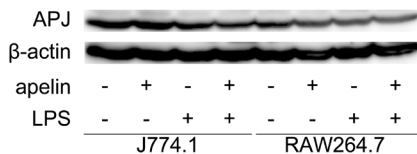


Figure 1. Expression of APJ was examined by Western blotting analysis in J774.1 and RAW264.7 cells. Cells were pretreated with or without [Pyr-1]-apelin-13 (1 μM) overnight and then stimulated with LPS (10 ng·mL⁻¹) for 6 h. Whole lysates were immunoblotted with anti-APJ antibody. The aliquots contained equivalent amounts of protein, which was verified by Western blotting for β-actin.

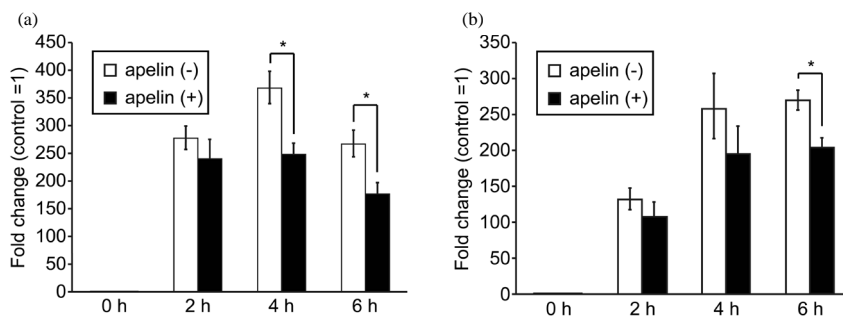


Figure 2. Effect of apelin on LPS-induced IL-1β and IL-6 mRNA expression in J774.1 cells. Cells were pretreated with or without [Pyr-1]-apelin-13 (1 μM) overnight and then stimulated with LPS (10 ng·mL⁻¹) for the indicated times. PCR amplification was performed using primers specific for IL-1β (a), IL-6 (b) and GAPDH. Values are expressed as fold change and compared with untreated control. Values shown represent the mean ± SD of results from triplicate cultures. **P* < 0.01, two-tailed Students *t*-test.

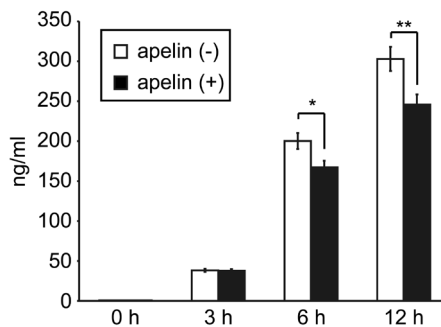


Figure 3. Effect of apelin on LPS-induced IL-6 protein secretion in J774.1 cells. Cells were pretreated with or without [Pyr-1]-apelin-13 (1 μM) overnight and then stimulated with LPS (10 ng·mL⁻¹) for the indicated times. The concentration of IL-6 in the supernatants of cultured J774.1 cells was analysed by ELISA. The data shown represent the mean ± SD of results from triplicate cultures. **P* < 0.05, ***P* < 0.01, two-tailed Students *t*-test.

sults strongly suggest that [Pyr1]-apelin-13 has anti-inflammatory properties by inhibiting IL-6 expression in macrophages.

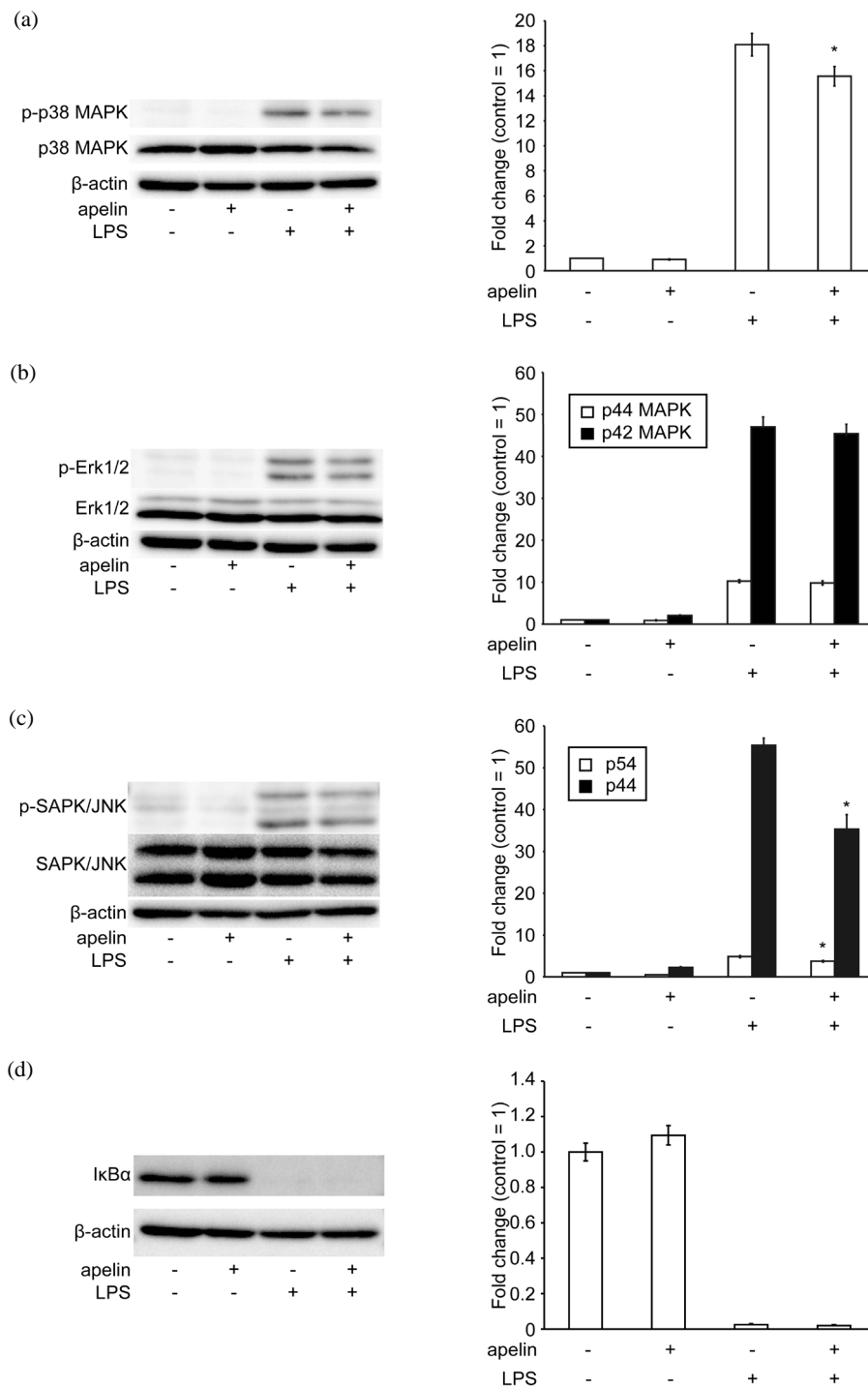


Figure 4. Effect of apelin on LPS induced MAPK phosphorylation in J774.1 cells. Cells were pretreated with or without [Pyr-1]-apelin-13 (1 μ M) overnight and then stimulated with LPS (10 ng/mL) for 30 min. Whole lysates were immunoblotted with anti-phospho-p38 MAPK, anti-p38 MAPK antibodies (a), anti-phospho-Erk1/2, anti-Erk1/2 antibodies (b), anti-phospho-SAPK/JNK and anti-SAPK/JNK antibodies (c), anti-I κ B α antibody (d). The aliquots used contained equivalent amounts of protein from the samples, which was verified by Western blotting for β -actin. The intensity of the band was quantified and calculated as the ratio of target protein to β -actin. The data shown represent the mean \pm SD of three independent experiments. * $P < 0.05$ (apelin-/LPS+ vs apelin+/LPS+), two-tailed Student's t -test.

3.3. Effect of Apelin on LPS-Induced Phosphorylation of p38 MAPK and JNK in J774.1 Cells

We examined whether [Pyr1]-apelin-13 inhibited LPS-stimulated intracellular signal transduction in J774.1 cells. Since LPS stimulation is known to activate MAPKs and NF- κ B, phosphorylation of MAPKs and degradation of I κ B (an endogenous inhibitor of NF- κ B) were observed by Western blotting analysis. As shown in **Figure 4**, pretreatment with [Pyr1]-apelin-13 significantly reduced p38 MAPK and JNK, but not ERK phosphorylation or degradation of I κ B in LPS-stimulated J774.1 cells. These data suggest that [Pyr1]-apelin-13 inhibited LPS-induced cell signalling in macrophages.

4. Discussion

In the present study, we demonstrated that pretreatment with [Pyr1]-apelin-13 diminished LPS-induced pro-inflammatory cytokine expression in the murine macrophage-like cell line, J774.1 cells. In addition, the phosphorylation of p38 MAPK and JNK induced by LPS decreased in [Pyr1]-apelin-13-treated J774.1 cells. To the best of our knowledge, this is the first report that the apelin peptide family has anti-inflammatory effects against bacterial LPS stimulation in macrophages.

Early studies on apelin suggested that it plays a role in the immune system. Apelin-36 and the more effective apelin-13 were found to suppress the production of interferon- γ , IL-2 and IL-4 in mouse spleen cells activated by CD3 cross-linking [15]. Since peripheral blood mononuclear cells treated with phytohaemagglutinin and IL-2 express APJ, the apelinergic system likely plays a role in immune activity [17]. In the present study, J774.1 and RAW 264.7 cells expressed APJ, suggesting that apelin has an immune function in macrophages. Previous studies also observed APJ expression in these two cell lines, which was consistent with our results [18]. Apelin administration has been reported to reduce levels of the 3T3-L1 adipocyte-driven pro-inflammatory adipokine acylation stimulating protein, which induces macrophage filtration in adipose tissue [19]. Apelin-13 promotes cholesterol efflux by enhancing the expression of ATP-binding cassette transporter A1 in foam cells, which suggests that apelin has an anti-atherogenic effect [20]. Considering these protective properties of apelin, overproduction of apelin in obese individuals may be an adaptive response before the onset of obesity-related disorders such as mild chronic inflammation, cardiovascular dysfunction and type 2 diabetes [21] [22].

While apelin pretreatment lowered LPS-induced IL-1 β and IL-6 production, we observed no significant effect of apelin alone on cytokine production in J774.1 cells. Previous studies, however, have demonstrated that apelin reduced IL-6 and MCP-1 production in J774A.1 cells [18]. Although different media or serum can affect the results, the reason for the discrepancy between the two reports remains unclear.

Toll-like receptors are large family members of type 1 transmembrane receptors that play an important role in the innate immune system. LPS activates the TLR4-mediated signalling pathway and leads to the activation of MAPKs to regulate the release of pro-inflammatory cytokines [5]. In addition, stimulation of murine macrophages by LPS induces the phosphorylation of ERK1/2, JNK and p38 MAPKs, which activates the expression of pro-inflammatory cytokines [23] [24]. In this study, we showed that apelin pretreatment suppressed LPS-induced JNK and p38 MAPK phosphorylation, suggesting that apelin has anti-inflammatory effects.

Adiponectin has been shown to possess anti-inflammatory properties against LPS stimulation in macrophages [25]-[27]. Both types of adiponectin isoforms, full-length (fAcrcp) and globular form (gAcrcp), suppressed LPS-induced MAPKs and NF κ B activation in macrophages. Although LPS-stimulated TNF- α mRNA transcription is directly suppressed by inactivation of NF- κ B and AP1 via the adiponectin receptor, transcription is also indirectly suppressed in a gAcrcp-induced IL-10-dependent manner at a later stage [27]. Recent studies demonstrated that the anti-inflammatory effects of gAcrcp mediate IL-10 and haeme oxygenase-1-dependent pathways [28], whereas the effects of fAcrcp are independent of IL-10 and are inducible by M2 polarisation in macrophages [25] [29]. In our experiments, apelin alone did not significantly increase IL-10 mRNA expression in J774.1 cells and RAW 264.7 cells (data not shown). Further experiments are needed to clarify that the molecular mechanisms of apelin in innate immunity remain unclear.

5. Conclusion

In conclusion, we demonstrated that [Pyr1]-apelin-13 has anti-inflammatory activity by inhibiting p38 MAPK and JNK, and reducing IL-1 β and IL-6 production in LPS-stimulated J774.1 cells.

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