

Antimicrobial Activity of Acidified Sodium Chlorite and Cell Free Culture Supernatant of Lactic Acid Bacteria against *Salmonella* Typhimurium

Sangeeta Singh^{1,2*}, Ajit Singh Yadav², Priyanka Bharti²

¹Applied Science Department, Indian Institute of Information Technology, Allahabad, India

²Food Microbiology Lab, Post Harvest Technology Division Central Avian Research Institute, Bareilly, India

Email: *sangeeta@iita.ac.in, sony134@gmail.com

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Abstract

Most methods used by food industries to decontaminate eggs involve washing of egg surface with various chemicals. In this study, the effectiveness of two organic decontaminants viz., acidified sodium chlorite (ASC) and cell free culture supernatant (CFCS) of two lactic acid bacteria (*Lactobacillus plantarum* and *Pediococcus cerevisiae*) was evaluated for the decontamination of spiked *Salmonella* Typhimurium on chicken egg shell surface. Acidified sodium chlorite at 100 µl/L concentration with the contact time of 20 min completely inhibited *S. Typhimurium* on egg shell surface while at 50 µl/L concentration 1 - 2 log₁₀ units reduction was observed in counts of *S. Typhimurium* as compared to control group. Likewise, CFCS of *P. cerevisiae* completely inhibited the growth of *S. Typhimurium* on 30 min contact, whereas *L. plantarum* and combination of both were revealed significant reduction in the counts of *S. Typhimurium* counts.

Keywords

Acidified Sodium Chlorite, Eggs, *Lactobacillus plantarum* and *Pediococcus cerevisiae* and *Salmonella* Typhimurium

1. Introduction

Eggs are highly nutritious and constitute an integral component of the human diet. They are also prone to con-

*Corresponding author.

tamination by various pathogenic microorganism viz. *E. coli* O157:H7, *Campylobacter jejuni*, *Yersinia enterocolitica*, etc. but among them *Salmonella* is the most predominant pathogen associated with it. Eggs and egg based products are the foods most frequently involved in *Salmonella* outbreaks and continue to be a significant public health problem. Although a wide range of *Salmonella* serovars can be isolated from eggs, *S. Typhimurium* and *S. Enteritidis* have been associated with most of the egg-borne outbreaks of salmonellosis [1]. Eggs get contaminated by *Salmonella* either from an infected hen (vertically) or during multiple steps along the food chain including processing, distribution, retail marketing and handling (horizontally) [2]. This calls for decontamination of *Salmonella* to reduce upward trend of incidence of egg-borne outbreaks and evolving appropriate methods or technology for the maintenance of egg quality during its movement from the farm to consumers.

Currently, most methods used by the food industry to decontaminate egg shell surfaces involve initially washing of them in detergent bath on conveyor rollers while being cleaned with brushes followed by treating them with various chemical solutions such as Lugol's solution, chlorihexadine, ethanol, and quaternary ammonium solutions. Exposure to ionized air, ozone in dry atmosphere, and pulsed UV-light has also been evaluated with varying degree of success [3] [4]. Even though, the agencies like Food Safety Inspection Service (FSIS), and Food Drug Administration (FDA) which are responsible for egg safety and the egg industries are investigating alternative decontamination techniques which could serve the public, minimize costs and benefit both the public and the industry [5].

Acidified sodium chlorite (ASC) has been approved by FDA for spray or dip application on various food products including poultry and poultry products, red meat, comminuted meat products, seafood and raw agricultural commodities [6]. ASC is highly effective broad spectrum antimicrobial produced by lowering the pH of a solution of sodium chlorite to 2.5 - 3.2 with any generally recognized safe acid. This combination results in the formation of the primary active component chlorous acid (HClO_2), which has strong antimicrobial properties primarily owing to its oxidative mode of action which is effective against bacteria, yeast, molds and viruses. A number of studies have described the strong efficacy of ASC on inactivation of pathogens like *E. coli* O157:H7 and *Salmonella* spp., [7] [8].

Lactic acid bacteria (LAB) are currently most investigated probiotic because they have received attention as a potential food preservative due to their strong antagonistic activity against many food-spoiling pathogenic organisms. Several lactic acid bacteria such as *Lactobacillus*, *Pediococcus*, *Bacteroides*, *Bifidobacterium*, *Enterococcus*, and *Helicobacter* have been used as probiotics alone or in combination [9]-[12]. The antimicrobial activity of lactic acid bacteria against pathogenic microorganisms involves numerous mechanisms including the production of metabolites with an antimicrobial action like organic acids (lactic acid), hydrogen peroxide and specific proteins complexes called bacteriocins [13]-[16]. The anti-*Salmonella* Typhimurium activity of LAB strains has been investigated by previous researchers [17]. The objective of this study was to compare the efficacy of ASC at various concentrations and cell free culture supernatant of two LAB strains (*Lactobacillus plantarum* and *Pediococcus cerevisiae*) in reducing the counts of artificially inoculated *S. Typhimurium* on chicken egg shell surface.

2. Materials and Methods

2.1. Bacterial Strains

Salmonella Typhimurium (E-2391) was procured from National Salmonella Centre (Veterinary), Indian Veterinary Research Institute, Izatnagar, Bareilly. *Lactobacillus plantarum* (NCDC-231) and *Pediococcus cerevisiae* (NCDC-252) were procured from National Dairy Research Institute, Karnal. For revitalization, freeze-dried bacteria were subcultured in Brain Heart Infusion (BHI, Himedia Laboratories, Mumbai) and De Man Rogosa Sharpe (MRS, Difco Laboratories, USA) broth and incubated at 37°C for 24 h. These strains were tested for their purity, morphological and biochemical characteristics and maintained by periodical subculturing on Nutrient agar (Himedia Laboratories, Mumbai) for *S. Typhimurium* and MRS agar (Difco Laboratories, USA) slants were used for *Lactobacillus plantarum* and *Pediococcus cerevisiae*.

2.2. Inoculum

Salmonella Typhimurium (E-2391) (Nalidixic acid resistant) was plated on Hektoen Enteric Agar (HEA, Himedia Laboratories, Mumbai). Single colony was transferred to BHI broth and incubated with agitation at 37°C for

24 h. *S. Typhimurium* cells were harvested by centrifugation ($4000 \times g$, 15 min) at 4°C . Pelleted cells in centrifuge tube were washed with 2 volumes of sterile normal saline solution (NSS, 0.85% NaCl) and resuspended in it. The bacterial growth was quantified as per method of ICMSF [18] and the cell density adjusted approximately to 10^6 cells/ml of bacterial suspension. Fresh inoculum was prepared for each experiment.

2.3. Cell Free Culture Supernatant (CFCS) of Lactic Acid Bacteria

Lactobacillus plantarum and *Pediococcus cerevisiae* were grown in MRS broth individually at 37°C for 24 h without shaking and CFCS from each culture was collected by centrifuging broth culture at $10,000 \times g$ for 30 min at 4°C . Centrifuged CFCS were passed through $0.22 \mu\text{m}$ Millipore filter (Himedia Laboratories, Mumbai) and stored at 4°C till used. pH of CFCS was determined and found to be 4.2 to 4.5.

2.4. Acidified Sodium Chlorite (ASC)

Sodium chlorite was acidified with citric acid following the manufacturer's (SRL, India) recommendations. Briefly, 1 g of citric acid and 0.8 g of sodium chlorite was dissolved in 10 ml distilled water and kept at room temperature for 30 min before use (stock solution). Different concentration of ASC viz. $50 \mu\text{l/L}$ (pH—3.0) and $100 \mu\text{l/L}$ (pH—2.71) were prepared in distilled water from the stock solution.

2.5. Inoculation of *S. Typhimurium* on Egg Shell Surface

Chicken eggs were obtained from commercial poultry farm and cleaned by washing in tap water to remove residual matter present on its outer surface followed by immersed in 70% ethanol for 10 min to ensure that eggs were free from any contamination and held at room temperature for half an hour before experiment. An area of $2 \times 1 \text{ cm}^2$ on the surface of each egg was inoculated with 0.1 ml of prepared inoculum of *S. Typhimurium* (10^6 cells/ml) and kept under laminar flow for 15 min before treatment to facilitate the attachment of *S. Typhimurium* cells to its surface.

2.6. Treatments of Inoculated Eggs with CFCS of Lactic Acid Bacteria and ASC

Inoculated eggs were treated with CFCSs of *Lactobacillus plantarum* and *Pediococcus cerevisiae* individually and in combination of both through dipping for different contact time (10 min, 20 min and 30 min). Sterile distilled water and MRS broth were employed as control for this experiment. Similarly, varying concentration ($50 \mu\text{l/L}$ and $100 \mu\text{l/L}$) of ASC was sprayed on spiked egg shell surface for a number of time intervals (5 min, 10 min, 20 min and 30 min) and sterile distilled water was used as control. Following exposure to above treatments, the marked area of egg shell was separated by cracking of egg, discarding the yolk and the white and crushed in sterile pestle-mortar containing 10 ml of normal saline solution, from which 10-fold serial dilution were prepared, plated on HE agar with 10% Nalidixic acid and incubated at 37°C for 24 h for enumeration of *S. Typhimurium* counts. Rappaport Vassiliadis (RV, Himedia Laboratories, Mumbai) and Tetrathionate (TT, Himedia Laboratories, Mumbai) broths were also inoculated from the NSS containing crushed egg shell and incubated at 37°C for 24 h for observing the growth of *S. Typhimurium*. Three trials were carried out for each experiment.

2.7. Statistical Analysis

Data obtained from above experiment was subjected to statistical analysis as per standard procedure of Snedecor and Cochran [19].

3. Results and Discussion

Acidified sodium chlorite was used for the decontamination of *S. Typhimurium* from egg shell surface. The mean log counts (\log_{10}) for *S. Typhimurium* on egg shell surface between different treatment groups and contact periods are presented in (Table 1). Different concentrations of ASC were found to be effective in reducing *S. Typhimurium* counts and significant differences ($p < 0.05$) were observed between treatment and control groups. Reduction in *S. Typhimurium* counts was directly proportional to concentration of ASC as well as contact time. Antimicrobial activity of ASC on spiked eggs was significantly influenced by the applied concentration with time. Thus, the reduction in *S. Typhimurium* counts increased with the increase in exposure time. Acidified

Table 1. Effect of ASC on decontamination of artificially inoculated *S. Typhimurium* (\log_{10} cfu/cm²) on egg shell surface.

S. No.	Contact time with ASC	<i>S. Typhimurium</i> count (\log_{10} cfu/cm ²)			Overall mean
		Group I	Group II	Group III	
1	5 min	3.71 ± 0.31	3.21 ± 0.32	3.25 ± 0.34	3.39 ^a ± 0.33
2	10 min	3.52 ± 0.34	3.18 ± 0.27	3.16 ± 0.34	3.28 ^a ± 0.34
3	15 min	3.46 ± 0.29	2.92 ± 0.29	2.29 ± 0.27	2.89 ^b ± 0.28
4	20 min	3.27 ± 0.27	2.71 ± 0.29	00 ± 0.29	1.91 ^c ± 0.29
	Overall mean	3.49 ^A ± 0.34	3.00 ^B ± 0.29	2.17 ^C ± 0.33	

Data are means ± SE of ten determinants. Means with superscripts a, b, c in a column and A, B in a row differ significantly ($p < 0.05$). Group I: Untreated control, Group II: 50 μ l/L ASC and Group III: 100 μ l/L ASC.

sodium chlorite at 100 μ l/L concentration with the contact time of 20 min completely inhibited *S. Typhimurium* on egg shell surface. Similar antimicrobial effect of ASC (0.1%) against *S. Typhimurium* on chicken skin has also been observed in an earlier study [20]. Other workers reported that ASC at 100 μ l/L concentration effectively reduced the bacterial growth on food products [7] [8]. Ruiz-Cruz *et al.*, (2007) reported that 100 μ l/L was the optimum concentration of ASC for maintaining overall quality, firmness, inhibiting microbial growth and prolonging the shelf life of food products. While, ASC with the concentration of 50 μ l/L indicated only reduction in *S. Typhimurium* counted instead of complete destruction. However, it reduced the counts by 1 - 2 log units as compared to control groups. Allende *et al.*, [21] and Ruiz-Cruz *et al.*, [8] also reported that lower concentration of ASC reduced the microbial population by 1 - 2 log cfu/g in fresh-cut cilantro and shredded carrots when compared with control group samples which were closely corroborative to our study.

In addition, enrichment (RV and TT broth) step was also performed to recover bacteria injured during the treatment. This enrichment was applied to determine whether the different concentration of ASC was able to eliminate the *S. Typhimurium* or merely suppress their growth. Among ASC treated samples, no viable cells of *S. Typhimurium* were recovered at the concentration of 100 μ l/L (20 min) indicating its bactericidal effect whereas ASC at lower concentrations had bacteriostatic effect on *S. Typhimurium* (Figure 1).

Conner (2001) and Caldwell *et al.* (2003) affirmed that ASC applied to inoculated fresh fruits and vegetables at 1.2 g/L for 1 min, killed at least 99.9% of *Salmonella* serotypes, *E. coli* O157:H7, and *L. monocytogenes* on carrots, strawberries, tomatoes, cucumbers, lettuce, cantaloupe and apples. Gonzalez *et al.*, (2004), Ruiz-Cruz *et al.*, (2007) and Inatsu *et al.*, (2005) [7] [8] [22] found a strong reduction in *E. coli* O157:H7, even under process water conditions, when using 0.5 and 1 g/L ASC on shredded carrots and Chinese cabbage. Lukasik *et al.*, 2003 [23] found that ASC at 0.1 and 0.2 g/L was more effective in reducing *E. coli* O157:H7 and *Salmonella* Montevideo populations on strawberries than stabilized chlorine dioxide or free chlorine disinfectants at comparable concentrations.

ASC is recognized as highly potent, broad spectrum antimicrobial system that has been successfully developed for uses in veterinary, food processing and medical device fields. ASC chemistry is principally due to chlorous acid which is the metastable oxychlorine species that forms on acidification of chlorite. Once formed, chlorous acid gradually decomposes to form chlorate ion, chlorine dioxide and chloride ion. Its antimicrobial efficacy derives from the uncharged chlorous acid, which is able to penetrate bacterial cell walls and disrupt protein synthesis by virtue of its reaction with sulfhydryl, sulfide and disulfide containing amino acids and nucleotides. The undissociated acid is thought to facilitate proton leakage into cells and thereby increase energy output of the cells to maintain their normal internal pH thereby also adversely affecting amino acid transport [24] [25].

Among Gram positive bacteria, lactic acid bacteria have been widely explored and considered to be a large reservoir of antimicrobial proteins that show a "natural effect" on food preservation [26]. Because of antimicrobial property possessed by lactic acid bacteria, cell free culture supernatant (CFCS) extracted from two different strains of lactic acid bacteria *viz.* *Lactobacillus plantarum* and *Pediococcus cerevisiae* individually and in combination were used in the present study for the decontamination of spiked *S. Typhimurium* on egg shell surface. Results indicated that CFCS of both the lactic acid bacterial strains on egg shell surface were found effective in reducing *S. Typhimurium* counts. Significant differences were observed between treatment groups as well as between varying contact periods of CFCS in reducing *S. Typhimurium* on egg shell surface. The mean

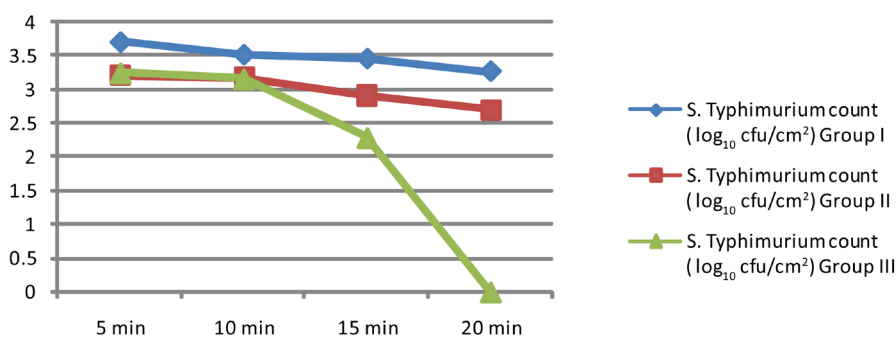


Figure 1. *Salmonella* Typhimurium count (\log_{10} cfu/cm²) on spiked egg shell surface treated with different concentration of ASC at various contact time.

logarithmic microbial counts (\log_{10}) for spiked *S. Typhimurium* on egg shell surface between different treatment group and control group are shown in **Table 2**. The counts of *S. Typhimurium* were more or less remained the same after 10 min exposure in all the groups and no reduction was observed among different treatment and control groups but slight reduction was observed after exposure of spiked eggs with CFCS for 20 min.

Means bearing different small letter superscripts (a, b, c) in a column and capital letter superscripts (A, B) in a row differ significantly ($p < 0.05$). LB: *Lactobacillus plantarum* extract, PD: *Pediococcus cerevisiae* extract, LB + PD: combination of *Lactobacillus plantarum* and *Pediococcus cerevisiae* extract & MRS: Fresh MRS broth without lactic acid bacteria inoculation.

In the present study CFCS of *P. cerevisiae* was found to be more effective and indicated the complete inhibition of spiked *S. Typhimurium* (10^6 cfu/ml) on the egg shell surface with contact time of 30 min. The complete inhibition of *S. Typhimurium* through *P. cerevisiae* may be due to the presence of pediocin (bacteriocin) or lower pH (4.5) effect exerted by its CFCS. Eijsink *et al.*, 1998 [27] conducted comparative study on antimicrobial effect of class IIa bacteriocins of lactic acid bacteria and reported that pediocin has broader inhibitory spectrum and our results are in agreement with this study.

The destructive activity of *L. plantarum* was produced mainly by non-lactic acid molecules that were present in CFCS of the *L. plantarum* [28]. But *Latobacillus plantarum* CFCS were not completely inhibited the spiked *S. Typhimurium* on egg shell surface. However, they reduced the counts by 1 - 2 \log_{10} units as compared to control groups ($p < 0.05$). Our results are in agreement with an earlier study where CFCS or bacteriocins produced by *Lactobacillus* were reported to have relatively narrow spectrum of antibacterial activity against important food borne pathogens like *Listeria monocytogenes*, *Yersinia enterocolitica*, *Campylobacter* spp. and *Salmonella* spp. [9]. It has been previously reported that *L. johnsonii* [29], *L. casi shirots* [30], *L. rhamnose* [31] and *L. acidophilus* [32] exerts antagonistic activity against Gram-negative pathogens.

Combination of CFCS of both *Lactobacillus plantarum* and *Pediococcus cerevisiae* also revealed incomplete inhibition but higher reduction was observed in comparison to *Lactobacillus plantarum* CFCS. *Lactobacillus plantarum* CFCS revealed only 1 \log_{10} reduction while CFCS of both reduced the *S. Typhimurium* counts upto 2 \log_{10} units as compared to control groups. The reason may be that the CFCS of *Pediococcus cerevisiae* complemented the antimicrobial activity of *Lactobacillus plantarum* CFCS by slightly lowering its pH. Moreover, longer contact time (30 min) with CFCS of *Lactobacillus plantarum* and *Pediococcus cerevisiae* were found to reduce spiked *S. Typhimurium* counts on egg shell surface significantly as compared to shorter contact periods of 10 and 20 min indicating the longer time requirement for CFCS to exhibit their antimicrobial effect. Enrichment in RV and TT broth was also performed for the samples treated with CFCS of *Lactobacillus plantarum* and *Pediococcus cerevisiae*. From all the treated samples viable cells of *S. Typhimurium* was recovered indicating that effect of CFCS was only bacteristatic not bactericidal (**Figure 2**).

The antimicrobial activity of probiotic lactic acid strains is known to be multifactorial. The efficacy of the spectrum of action of lactic acid bacteria against pathogenic microorganisms is determined by the production of metabolites with an antimicrobial action, including organic acids such as lactic acid and acetic acid [13], hydrogen peroxide which inhibits the growth of pathogens based on a strong oxidant effect on bacterial cells [14] or molecular destruction of nucleic acid and cell proteins and the production of specific proteins or protein complexes called bacteriocins. The mechanism of the antimicrobial activity of probiotic lactic acid bacteria strains

Table 2. Effect of extracts of lactic acid bacteria on decontamination of artificially inoculated *S. Typhimurium* (\log_{10} cfu/cm²) on egg shell surface.

S. No.	Treatment	<i>S. Typhimurium</i> count (\log_{10} cfu/cm ²)			Overall mean
		10 min	20 min	30 min	
1	LB	3.38 ± 0.29	3.2 ± 0.27	2.45 ± 0.29	3.19 ^{ab} ± 0.29
2	PD	3.34 ± 0.28	2.02 ± 0.31	00 ± 0.28	2.12 ^c ± 0.29
3	LB+PD	3.46 ± 0.29	2.77 ± 0.29	2.04 ± 0.34	2.76 ^b ± 0.30
4	MRS	3.42 ± 0.29	3.43 ± 0.26	3.48 ± 0.29	3.44 ^a ± 0.29
	Overall mean	3.43 ^A ± 0.29	3.22 ^A ± 0.28	2.39 ^B ± 0.29	

Means bearing different small letter superscripts (a, b, c) in a column and capital letter superscripts (A, B) in a row differ significantly ($p < 0.05$). LB: *Lactobacillus plantarum* extract, PD: *Pediococcus cerevisiae* extract, LB + PD: combination of *Lactobacillus plantarum* and *Pediococcus cerevisiae* extract & MRS: Fresh MRS broth without lactic acid bacteria inoculation.

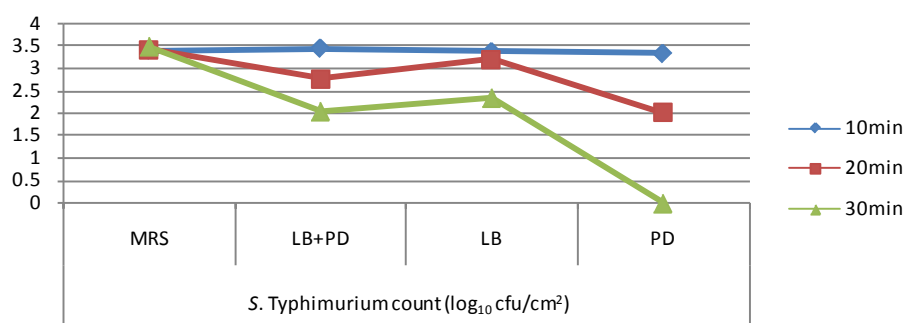


Figure 2. *Salmonella Typhimurium* count (\log_{10} cfu/cm²) on spiked egg shell surface treated with different extracts of lactic acid bacteria at various contact time. (LB: *Lactobacillus plantarum* extract, *Pediococcus cerevisiae* extract, LB + PD: combination of *Lactobacillus plantarum* and *Pediococcus cerevisiae* extract and MRS: Fresh MRS broth without lactic acid bacteria inoculation).

that leads to killing of bacterial pathogens may be due to synergistic action of lactic acid and non-lactic acid molecules [33].

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References

- [1] EFSA (2009) The Community Summary Report on Food-Borne Outbreaks in the European Union in 2007. *EFSA Journal*, **271**, 1-102.
- [2] Jones, F.T., Rive, D.V. and Carey, J.B. (1995) Salmonella Contamination in Commercial Eggs and an Egg Production Facility. *Poultry Science*, **74**, 753-757. <http://dx.doi.org/10.3382/ps.0740753>
- [3] Davies, R.H. and Breslin, M. (2003) Investigations into Possible Alternative Decontamination Methods for Salmonella Enteritidis on the Surface of Table Eggs. *Journal of Veterinary Medicine B*, **50**, 38-41. <http://dx.doi.org/10.1046/j.1439-0450.2003.00622.x>
- [4] Hierro, E., Manzano, S., Ordonez, J.A., Hoz, L. and Fernandez, M. (2009) Inactivation of Salmonella Enterica Serovar Enteritidis on Shell Eggs by Pulsed Light Technology. *International Journal of Food Microbiology*, **135**, 125-130. <http://dx.doi.org/10.1016/j.ijfoodmicro.2009.07.034>
- [5] CFSAN (2000) U.S. Egg Safety Action Plan. FDA, Rockville. <http://www.cfsan.fda.gov/~dms/fs-eggs3.html>
- [6] Food and Drug Administration (FDA) (2000) Acidified Sodium Chlorite Solutions. Code of Federal Regulations, 21CFR173.325. <http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm>

- [7] Gonzalez, R.J., Luo, Y., Ruiz-Cruz, S. and Mcevoy, J.L. (2004) Efficacy of Sanitizers to Inactivate *Escherichia coli* O157:H7 on Fresh-Cut Carrot Shreds under Simulated Process Water Conditions. *Journal of Food Protection*, **67**, 2375-2380.
- [8] Ruiz-Cruz, S., Acedo-Félix, E., Díaz-Cinco, M., Islas-Osuna, M.A. and González-Aguilar, G.A. (2007) Efficacy of Sanitizers in Reducing *Escherichia coli* O157:H7, *Salmonella* spp. and *Listeria monocytogenes* Populations on Fresh-Cut Carrots. *Food Control*, **18**, 1383-1390. <http://dx.doi.org/10.1016/j.foodcont.2006.09.008>
- [9] Nowroozi, J., Mirzaii, M. and Norauzi, M. (2004) Study of Lactobacillus as Probiotic Bacteria. *Iranian Journal of Public Health*, **33**, 1-7.
- [10] Lievin, V., Peiffer, I., Hudault, S., Rochat, F., Brassart, D., Neeser, J.R. and Servin, A.L. (2000) Bifidobacterium Strains from Resident Infant Human Gastrointestinal Microflora Exert Antimicrobial Activity. *Gut*, **47**, 646-652. <http://dx.doi.org/10.1136/gut.47.5.646>
- [11] Yoon, M.Y., Lee, K. and Yoon, S.S. (2014) Protective Role of Gut Commensal Microbes against Intestinal Infections. *Journal of Microbiology*, **52**, 983-989. <http://dx.doi.org/10.1007/s12275-014-4655-2>
- [12] Kim, J.E., Kim, M.S., Yoon, Y.S., Chung, M.J. and Yum, D.Y. (2014) Use of Selected Lactic Acid Bacteria in the Eradication of *Helicobacter pylori* Infection. *Journal of Microbiology*, **52**, 955-962. <http://dx.doi.org/10.1007/s12275-014-4355-y>
- [13] Gilliland, S.E. and Speck, M.L. (1972) Interactions of Food Starter Cultures and Food Borne Pathogens: Lactic Acid Streptococci versus Staphylococci and Salmonellae. *Journal of Milk Food and Technology*, **35**, 307-310.
- [14] Price, R.J. and Lee, J.S. (1970) Inhibition of *Pseudomonas* spp. by Hydrogen Peroxide Producing. *Lactobacilli Milk and Food Technology*, **33**, 3-18.
- [15] Servin, A.L. (2004) Antagonistic Activities of Lactobacilli and Bifidobacteria against Microbial Pathogens. *FEMS Microbiology Review*, **28**, 405-440. <http://dx.doi.org/10.1016/j.femsre.2004.01.003>
- [16] Reid, G., Jass, J., Sebulsky, M.T. and McCormick, J.K. (2003) Potential Use of Probiotics in Clinical Practice. *Clinical Microbiology Review*, **16**, 658-672. <http://dx.doi.org/10.1128/CMR.16.4.658-672.2003>
- [17] Coconnier-Polter, M.H., Levin, V. and Servin, A.L. (2005) A *Lactobacillus acidophilus* Strain of Human Gastrointestinal Microbiota Origin Elicits Killing of Enterovirulent *Salmonella enterica* Serovar Typhimurium by Triggering Lethal Bacterial Membrane Damage. *Applied Environmental Microbiology*, **71**, 6115-6120. <http://dx.doi.org/10.1128/AEM.71.10.6115-6120.2005>
- [18] ICMSF (1978) International Commission for Microbiological Specialists for Food. Micro-Organism in Food-1, Their Significance and Methods of Enumeration. 2nd Edition, University of Toronto Press, Toronto, 115-118.
- [19] Snedecor, G.W. and Cochran, W.G. (1980) Statistical Methods. 7th Edition, Oxford and IBH Publishing Co., Calcutta.
- [20] Haydar, O. and Sebnem, P. (2006) Acidified Sodium Chlorite, Trisodium Phosphate and Populations of *S. Typhimurium* and *Staphylococcus aureus* on Chickens-Breast Skin. *Journal of Food Processing and Preservation*, **30**, 237-247.
- [21] Allende, A., Mcevoy, J., Tao, Y. and Luo, Y. (2009) Antimicrobial Effect of Acidified Sodium Chlorite, Sodium Chlorite, Sodium Hypochlorite, and Citric Acid on *Escherichia coli* O157:H7 and Natural Microflora of Fresh-Cut Cilantro. *Food Control*, **20**, 230-234. <http://dx.doi.org/10.1016/j.foodcont.2008.05.009>
- [22] Inatsu, Y., Bari, M.L., Kawasaki, S., Isshiki, K. and Kawamoto, S. (2005) Efficacy of Acidified Sodium Chlorite Treatments in Reducing *Escherichia coli* O157:H7 on Chinese Cabbage. *Journal of Food Protection*, **68**, 251-255.
- [23] Lukasik, J., Bradley, M.L., Scott, T.M., Dea, M., Koo, A., Hsu, W.Y., *et al.* (2003) Reduction of Poliovirus 1, Bacteriophages, *Salmonella* Montevideo, and *Escherichia coli* O157:H7 on Strawberries by Physical and Disinfectant Washes. *Journal of Food Protection*, **66**, 188-193.
- [24] Warf, C.C. (2001) The Chemistry and Mode of Action of Acidified Sodium Chlorite. 2001 *IFT Annual Meeting*, New Orleans, 23-27 June 2001, 1-91.
- [25] Martin, R.W. (2012) Method and Composition for *In-Situ* Generation of Chlorous Acid. US Patent No. 20120107418.
- [26] Cleveland, J., Montville, J.J., Nes, I.F. and Chikindas, M. (2001) Bacteriocins: Safe, Natural Antimicrobials for Food Preservation. *International Journal of Food Microbiology*, **71**, 1-20. [http://dx.doi.org/10.1016/S0168-1605\(01\)00560-8](http://dx.doi.org/10.1016/S0168-1605(01)00560-8)
- [27] Eijsink, V.G.H., Skeie, M., Middelhoven, P.N., Brurberg, B.M. and Nes, F.I. (1998) Comparative Studies of Class IIa Bacteriocins of Lactic Acid Bacteria. *Applied Environmental Microbiology*, **64**, 3275-3281.
- [28] Fayol-Messaoudi, D., Coconnier-Polter, M.H., Lievin, L.E., Moal, V., Atasi, F., Berger, C.N. and Servin, A.L. (2007) The *L. plantarum* Strain ACA-DC28 Isolated from a Greek Cheese Demonstrates Antagonistic Activity *in Vitro* and *in Vivo* against *Salmonella enterica* Serovar Typhimurium. *Journal of Applied Microbiology*, **103**, 647-655. <http://dx.doi.org/10.1111/j.1365-2672.2007.03293.x>
- [29] Tuomola, E.M., Ouwchand, A.C. and Salminen, S.J. (1999) The Effect of Probiotic Bacteria on the Adhesion of Pathogens to Human Intestinal Mucosa. *FEMS Immunology and Medical Microbiology*, **26**, 137-142.

<http://dx.doi.org/10.1111/j.1574-695X.1999.tb01381.x>

- [30] Lee, Y.K., Puong, K.Y., Ouwehand, A.C. and Salminen, S. (2003) Displacement of Bacterial Pathogens from Mucus and Caco 2 Cells Surface by Lactobacilli. *Journal of Medical Microbiology*, **52**, 925-930. <http://dx.doi.org/10.1099/jmm.0.05009-0>
- [31] McGroarty, J.A. and Reid, G. (1998) Detection of Lactobacillus Substances That Inhibits *E. coli*. *Canadian Journal of Microbiology*, **34**, 974-978. <http://dx.doi.org/10.1139/m88-171>
- [32] Chen, X., Xu, J., Shuai, J., Che, J., Zhang, Z. and Fang, W. (2007) The S-Layer Proteins of *Lactobacillus crispatus* Strain ZT001 Is Responsible for Competitive Exclusion against *E. coli* O157:H7 and *S. Typhimurium*. *International Journal of Food Microbiology*, **115**, 307-312. <http://dx.doi.org/10.1016/j.ijfoodmicro.2006.11.007>
- [33] Fayol-Messaoudi, D., Berger, C.N., Coconneir-Polter, M.H., Levin, V. and Servin, A.L. (2005) pH-, Lactic Acid- and Non-Lactic Acid-Dependent Activities of Probiotic Lactobacilli against *Salmonella enterica* Serovar Typhimurium. *Applied Environmental Microbiology*, **71**, 6008-6013. <http://dx.doi.org/10.1128/AEM.71.10.6008-6013.2005>