

Essential Oil of *Phyllanthus reticulatus* Poiret from Nigeria*

Akintayo L. Ogundajo¹, Afusat M. Aruna¹, Ayo O. Owolabi¹, Isiaka A. Ogunwande^{1#}, Guido Flamini²

¹Natural Products Research Unit, Department of Chemistry, Faculty of Science, Lagos State University, PMB 0001, LASU Post Office, Ojo, Lagos, Nigeria

²Dipartimento di Scienze Farmaceutiche, sede Chimica Bioorganica e Biofarmacia, Universita di Pisa, Via Bonanno 33, Pisa, Italy
Email: #isiaka.ogunwande@lasu.edu.ng

Received 13 October 2014; revised 20 November 2014; accepted 21 December 2014

Copyright © 2015 by authors and Scientific Research Publishing Inc.
This work is licensed under the Creative Commons Attribution International License (CC BY).
<http://creativecommons.org/licenses/by/4.0/>



Open Access

Abstract

The chemical composition of the essential oil obtained by hydrodistillation from the leaves of *Phyllanthus reticulatus* Poiret (Euphorbiaceae) growing in Nigeria has been studied. The constituents of the oil were analyzed by means of gas chromatography (GC-FID) and gas chromatography coupled with mass spectrometry (GC-MS). Monoterpenes (64.9%) were the dominant class of compounds, followed by sesquiterpenes (23.0%). The major constituents were α -pinene (6.0%), sabinene (7.6%), β -pinene (18.1%), linalool (6.9%) and camphor (7.7%), among the monoterpenes, and β -caryophyllene (11.9%) and germacrene D (8.6%) among the sesquiterpenes. This is the first report on the volatile constituents of *Phyllanthus reticulatus*.

Keywords

Phyllanthus reticulatus, Essential Oil Composition, Monoterpenes, Sesquiterpenes

1. Introduction

Phyllanthus reticulatus Poiret., (Family Euphorbiaceae) is a many branched deciduous shrub or small tree sometimes partially scrambling, usually 1 - 5 m high. *P. reticulatus* usually has a distinct smell that is emitted by the minute flowers when they open towards the early evening. The bark is light reddish-brown or grey-brown with

*Presented as Poster (BV-55) at the 44th International Symposium on Essential Oil (ISEO-44), Danubius Hotel, Flamenco, Budapest, Hungary, September 8-12, 2013.

#Corresponding author.

hairy stems when young, which become smooth with age. The leaves alternate along slender branches. They are up to 25 cm long and appear as leaflets of large pinnate leaves. The leaves are thinly textured, usually hairless. It flowers from September to October, but the flowering season can extend from July onwards. *P. reticulatus* has very small, roundish berry like fruits that are green at first, turning purple-black, 4 - 6 mm in diameter [1]. Extracts of the plant is known to possess both analgesic and anti-inflammatory activities [2]-[4]. Aqueous extract of *P. reticulatus* can be utilized for prevention of atherosclerosis in hypercholesterolemic patients [5]. There are reports which describe the antiviral [6], antibacterial [7], hepatoprotective [8], antioxidant [9], potential RNase H inhibition and protection against the viral cytopathic effects of HIV-1 [10], antidiabetic [11] and hypoglycemic [12] activities.

Some biologically active compound such as 2- α -hydroxyfriedel-4(23)-en-3-one and other triterpenoids [13], purine, 3-(3-methylbut-2-en-1-yl)isoguanine and cleistanthane-type diterpenoid glucoside, 19-hydroxyspruceanol 19-*O*- β -D-glucopyranoside [14], (5R, 6R)-4,6-Dimethoxycarbonyl-5-[2',3',4'-trihydroxy-6'-(methoxycarbonyl)phenyl]-5,6-dihydro-2H-pyran-2-one, 3,4,3'-tri-*O*-methylsuccinic acid and methyl gallate [15], reticulatusides A and B [16], cytotoxic scopoletin [17] and flavonoid glycosides [18] [19] have been isolated from this plant. The isolation of lupeol, stigmaterol and lupeol acetate from the plant have been reported [20]. Regarding the volatile constituents, there appears to be no published work.

The objective of the present work was to examine the volatile constituents of this plant for future exploration. Our finding into the volatile oils of some poorly studied Nigerian flora was recently published [21].

2. Materials and Methods

2.1. Plant Sample

Leaves of *P. reticulatus* were collected from a location in Ibe fun, Odogbolu, Ogun State, Nigeria, on March, 2012. Botanical identification was performed by Messrs Ugbogu O.A and Shasanya, O.S., at the Herbarium Headquarters, Forestry Research Institute of Nigeria (FRIN), Ibadan, Nigeria, where voucher specimen (FHI 109586) has been deposited for future reference.

2.2. Extraction of Essential Oil

Aliquots (400 g) of the air-dried and pulverized plant sample were subjected to hydrodistillation for 3 h using a Clevenger-type apparatus in accordance with the British Pharmacopoeia specification [22] to produce a pale yellow essential oil.

2.3. Analysis of the Oil

GC analysis was accomplished with a HP-5890 Series II instrument equipped with a HPWax and HP-5 capillary columns (both 30 m \times 0.25 mm, 0.25 μ m film thickness), working with the following temperature program: 60°C for 10 min, rising at 5°C/min to 220°C. The injector and detector temperatures were maintained at 250°C; carrier gas nitrogen (2 mL/min); detector dual, FID; split ratio 1:30. The volume injected was 0.5 μ L. The relative proportions of the oil constituents were percentages obtained by FID peak-area normalization without the use of response factor.

GC-EIMS analysis was performed with a Varian CP-3800 gas-chromatograph equipped with a HP-5 capillary column (30 m \times 0.25 mm; film thickness 0.25 μ m) and a Varian Saturn 2000 ion trap mass detector. Analytical conditions: injector and transfer line temperature 220°C and 240°C respectively; oven temperature programmed from 60°C - 240°C at 3°C/min; carrier gas was helium at a flow rate of 1 mL/min; injection of 0.2 μ L (10% hexane solution); split ratio 1:30. Mass spectra were recorded at 70 eV. The acquisition mass range was 30 - 300 m/z at a scan rate of 1 scan/sec.

2.4. Identification of the Constituents

Identification of the constituents was based on comparison of the retention times with those of authentic samples, comparing their linear retention indices (LRI) relative to the series of n-hydrocarbons, and on computer matching against commercially available spectral [23]-[25]. Further identifications were also made possible by the use of homemade library mass spectra built up from pure substances and components of known oils and MS literature data. Moreover, the molecular weights of all the identified substances were confirmed by GC-CIMS, using

MeOH as CI ionizing gas.

3. Results and Discussion

P. reticulatus yielded low content of essential oil 0.12% (v/w) on a dry weight basis. **Table 1** shows the identities of 84 compounds identified in the oil of *P. reticulatus*, accounting for 99.7% of the total oil contents. **Figure 1** depicts the GC chromatogram of the essential oil. The classes of compounds identified in the oil were monoterpene hydrocarbons (42.0%), oxygenated monoterpenoids (23.5%), sesquiterpene hydrocarbons (29.4%), oxygenated sesquiterpenoids (3.6%) and non-terpene derivatives (1.2%). The major oil constituents were α -pinene (6.0%), sabinene (7.6%), β -pinene (18.1%), linalool (6.9%) and camphor (7.7%), among the monoterpenes; and β -caryophyllene (11.9%) and germacrene D (8.6%) among the sesquiterpenes. This may represent the first analysis of the oil of this species.

Regardless of *Phyllanthus* being large family, with about 1000 species, the essential oils of *P. reticulatus* has not been investigated. However, the volatile constituents of few species grown have been reported. Phytol (21.5%), β -citronellol (17.7%), *trans*-geraniol (13.5%), *cis*-3-hexenol (12.6%) and 1-hexanol (11.3%) were the major constituents of *Phyllanthus salviaefolius* H.B.K. [26]. However, linalool (36.4%) and phytol (13.0%) dominated the oil of *Phyllanthus amarus* Sch. and Thonn [27]. Volatile compounds have been isolated from *P. acidus* (L.) Skeels fruits fermented for 1, 3 and 6 months. Among the 46 compounds identified, acids and alcohols dominated the volatiles profile; acids particularly characterized the quantitative profile of the volatile compounds after 6 months of fermentation. Other significant changes were in the sesquiterpenes, with increments of δ - and α -cadinene after 3 months of fermentation, and α -cadinol and τ -muurolol after 6 months [28]. *Phyllanthus arenarius* Beille in Lecomte has *n*-hexadecanoic acid (14.0%), 1,2-benzene dicarboxylic acid, bis (2-methylpropyl) ester (12.7%) and di-*n*-octyl phthalate (10.3%) as its main compounds [29] while *Phyllanthus urinata* L., was rich in 3,3,5-trimethylcyclohexanone (17.2%) and *n*-hexadecanoic acid (12.4%). The abundant of 3,3,5-trimethylcyclohexanone (12.4%) and 3,7-dimethyl-1,6-octadien-3-ol (10.2%) was reported in the oil of *Phyllanthus niriru* L. [29]. The essential oil of *Phyllanthus emblica* L. contained high amounts of β -caryophyllene, β -bourbonene, 1-octen-3-ol, thymol, and methyleugenol [30]. Another investigation reported that β -bourbonene, heptadecanol, pentadecanone, thymol, β -caryophyllene, β -neoclovene, nerol and borneol were the major compounds were the main oil contents of the plant [31]. (*E*)-Isoelemicin (36.40%) was the main compounds of *Phyllanthus muellerianus* (Kuntze) Excel which also showed antimicrobial property [32].

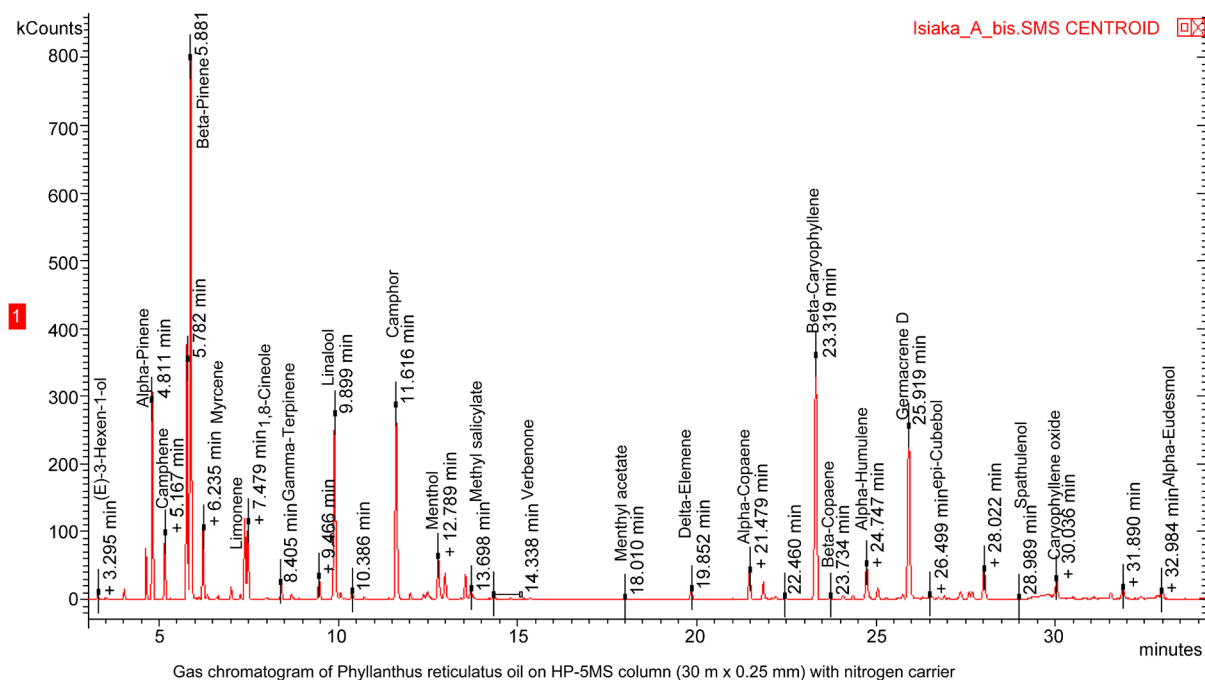


Figure 1. Gas chromatogram of *Phyllanthus reticulatus* oil on HP-5MS column (30 m x 0.25 mm) with nitrogen carrier.

Table 1. Essential oil constituents of *Phyllanthus reticulatus*.

Compounds ^a	Retention Times ^b	RI ^c	RI ^d	Percentage (%)
(<i>E</i>)-Hex-3-en-1-ol	3.29	854	850	0.4
Hexan-1-ol	3.48	872	863	Tr
<i>n</i> -Nonane	4.02	900	900	0.3
Tricyclene	4.55	928	921	Tr
α -Thujene	4.64	931	924	1.3
α -Pinene	4.81	940	932	6.0
Camphene	5.16	954	946	1.9
Thuja-2,4(10)-diene	5.29	958	953	Tr
Benzaldehyede	5.45	962	962	Tr
Sabinene	5.78	977	969	7.6
β -Pinene	5.88	980	974	18.1
Octan-3-one	6.11	988	979	Tr
Myrcene	6.23	992	988	2.4
Octan-3-ol	6.34	995	988	Tr
α -Phellandrene	6.64	1005	1002	Tr
α -Terpinene	7.01	1018	1014	0.4
<i>p</i> -Cymene	7.26	1027	1020	0.2
Limonene	7.39	1031	1024	2.6
1,8-Cineole	7.48	1034	1026	2.7
(<i>Z</i>)- β -Ocimene	7.69	1041	1032	Tr
(<i>Z</i>)-Oct-3-en-1-ol	7.97	1047	1047	Tr
(<i>E</i>)- β -Ocimene	8.04	1051	1044	Tr
γ -Terpinene	8.41	1062	1054	0.6
<i>cis</i> -Sabinene hydrate	8.70	1070	1065	0.2
<i>cis</i> -Linalool oxide (furanoid)	8.89	1075	1067	Tr
Non-1-en-3-ol	9.16	1083	1088	Tr
Terpinolene	9.47	1089	1086	0.9
Linalool	9.90	1099	1095	6.9
<i>n</i> -Nonanal	10.07	1103	1100	0.2
Octen-1-ol acetate	10.39	1107	1110	0.3
<i>cis-p</i> -Menth-2-en-1-ol	10.72	1122	1118	Tr
<i>trans-p</i> -Menth-2-en-1-ol	11.43	1125	1136	Tr
Camphor	11.62	1144	1141	7.7
Menthone	12.01	1155	1158	0.3
Pinocarvone	12.37	1163	1160	0.2

Continued

Borneol	12.49	1166	1165	0.5
Menthol	12.79	1174	1167	1.9
Terpinen-4-ol	12.97	1178	1174	1.1
(Z)-Hex-3-enyl butanoate	13.41	1186	1184	Tr
α -Terpineol	13.54	1190	1186	1.0
Methyl salicylate	13.70	1192	1190	0.6
<i>n</i> -Decanal	14.21	1205	12.01	Tr
Verbenone	14.34	1208	1204	0.3
β -Cyclocitral	14.80	1218	1217	Tr
Nerol	15.18	1228	1227	Tr
3-Methyl-hex-3-en-1-yl butanoate	15.36	1236	1232	Tr
Piperitone	16.24	1253	1249	Tr
Menthyl acetate	18.01	1294	1294	0.1
δ -Elemene	19.85	1340	1335	0.5
α -Cubebene	20.36	1351	1345	Tr
Eugenol	20.74	1358	1356	Tr
α -Copaene	21.48	1376	1374	1.5
β -Bourbonene	21.58	1384	1387	1.0
β -Cubebene	22.11	1390	1387	Tr
β -Elemene	22.28	1392	1389	0.2
Cyperene	22.46	1398	1398	0.2
Isocaryophyllene	22.79	1405	1408	Tr
Dodecanal	22.97	1408	1408	Tr
β -Caryophyllene	23.32	1418	1417	11.9
β -Copaene	23.73	1429	1430	0.2
α -Guaiene	24.08	1439	1437	0.2
Aromadendrene	24.35	1441	1439	0.2
α -Humulene	24.75	1455	1452	1.8
<i>allo</i> -Aromadendrene	25.05	1461	1458	0.5
<i>cis</i> -Muurolo-4(14),5-diene	25.29	1463	1465	Tr
γ -Muurolole	25.74	1477	1479	1.8
Germacrene D	25.92	1480	1484	8.6
(<i>E</i>)- β -Ionone	26.18	1485	1487	Tr
<i>cis</i> - β -Guaiene	26.31	1490	1492	Tr
<i>epi</i> -Cubebol	26.50	1494	1493	0.4

Continued

<i>trans-β</i> -Guaiene	26.75	1500	1502	Tr
<i>α</i> -Bulnesene	26.90	1505	1509	0.3
<i>δ</i> -Amorphene	27.02	1512	1511	Tr
Cubebol	27.35	1515	1514	0.5
<i>δ</i> -Cadinene	27.69	1524	1522	0.4
Germacrene B	28.83	1556	1559	0.1
Spathulenol	28.99	1576	1577	0.4
Caryophyllene oxide	30.04	1581	1582	1.2
Humulene epoxide II	31.07	1606	1608	Tr
1,10-di- <i>epi</i> -Cubenol	31.54	1614	1618	0.3
<i>τ</i> -cadinol	32.39	1641	1639	0.2
Cubenol	32.56	1647	1645	Tr
<i>α</i> -Cadinol	32.86	1652	1652	0.2
<i>α</i> -Eudesmol	32.98	1654	1652	0.4
TOTAL				99.7
Monoterpene hydrocarbons				42.0
Oxygenated monoterpenes				23.5
Sesquiterpene hydrocarbons				29.4
Oxygenated sesquiterpenes				3.6
Others				1.2

^aElution order on HP-5MS column; ^bRetention time in order with respect to the chromatogram (Figure 1); ^cRetention indices on HP-5 MS capillary column; ^dLiterature retention indices (References 23 - 25); Tr, Trace amounts < 0.1%.

The main compounds of *P. salviaefolius* could not be identified in *P. reticulatus*. Except for linalool, the quantitatively significant compounds of *P. amarus* were conspicuously absent from *P. reticulatus*. Also, the low contents of *δ*-cadinene and *α*-cadinol, and the absence of the sesquiterpenoid compounds such as *α*-cadinene and *τ*-muurolol in this study makes the composition differ from that found in *P. acidus*. Also, the major compounds in the oils of *P. arenarius*, *P. urinate* and *P. niriru* were absent in *P. reticulatus*. In addition, the oil of *P. reticulatus* could be distinguished from those of *P. embelica* by its lack of thymol, methyleugenol, heptadecanol, pentadecanone and nerol. Notably, (*E*)-isoelemicin, the main compound of *P. muellerianus* was not identified in *P. reticulatus*.

4. Conclusion

The chemical constituents of essential oil obtained from *P. reticulatus* grown in Nigeria are being reported for the first time. In addition, a comparison of the chemical composition was made with the other known essential oils from *Phyllanthus* plants. It could be seen that the essential oils of *Phyllanthus* plants exhibit high chemical variability. Each species has its own compositional pattern different from other. The very high content of compounds identified in the oil of *P. reticulatus* may be an important chemical and economic characteristic of the oil sample.

Acknowledgements

Authors are grateful to Messrs Ugbogu and Shasanya for the botanical identification of the plant.

References

- [1] Palgrave, M.C. (2002) Trees of Southern Africa. Third Edition, Struik Publisher, Cape Town.
- [2] Saha, A., Masud, M.A., Bachar, S.C., Kundu, J.K., Nahar, L., Datta, B.K. and Sarker, S.D. (2007) The Analgesic and Anti-inflammatory Activities of The Extracts of *Phyllanthus reticulatus* in Mice Model. *Pharmaceutical Biology*, **45**, 355-359. <http://dx.doi.org/10.1080/13880200701212973>
- [3] Sunil, K., Shalini, S., Dinesh, K., Tarun, K., Renu, A. and Karunesh, A. (2012) Pharmacognostic Study and Anti-inflammatory Activity of *Phyllanthus reticulatus* Poir. Fruit. *Asian Pacific Journal of Tropical Disease*, **2**, S332-S335. [http://dx.doi.org/10.1016/S2222-1808\(12\)60177-7](http://dx.doi.org/10.1016/S2222-1808(12)60177-7)
- [4] Patel, J. and Gaudani, K. (2011) Evaluation of Anti-Inflammatory Activity of Petroleum Ether and Methanolic Extract of *Phyllanthus reticulatus* Leaves. *Journal of Advanced Pharmacy Education and Research*, **1**, 266-270.
- [5] Maruthappan, V. and Shree, K.S. (2010) Effects of *Phyllanthus reticulatus* on Lipid Profile and Oxidative Stress in Hypercholesterolemic Albino Rats. *Indian Journal of Pharmacology*, **42**, 388-391. <http://dx.doi.org/10.4103/0253-7613.71923>
- [6] Das, B.K., Shohel, M., Pavel, A.M., Akhter, N., Yasmin, T., Bhattacharjee, R., Das, B. and Hannan, J.M.A. (2011) Anti-Hepatitis B Viral Activity of *Phyllanthus reticulatus*. *Bangladesh Pharmaceutical Journal*, **14**, 11-14.
- [7] Akhtar, M.A., Sharmin, S.A., Rahman, M.A., Rahman, M.M., Khalekuzzaman, M.K., Anisuzzaman, M. and Alam, M.F. (2013) Antibacterial Potential of Leaf Extracts from *Phyllanthus reticulatus*. *International Journal of Pharmacy and Research Development*, **5**, 85-95.
- [8] Das, B.K., Bepary, S., Datta, B.K., Choudahry, A.K.A., Ali, M.S. and Rouf, A.S.S. (2008) Hepatoprotective Activity of *Phyllanthus reticulatus*. *Pakistan Journal of Pharmaceutical Science*, **21**, 333-337.
- [9] Sani, M.F., Kouhsari, S.M. and Moradabadi, L. (2012) Effects of Three Medicinal Plants Extracts in Experimental Diabetes: Antioxidant Enzymes Activities and Plasma Lipids Profiles in Comparison with Metformin. *Iranian Journal of Pharmaceutical Research*, **1**, 897-903.
- [10] Tai, B.H., Nhut, N.D., Nhiem, N.X., Quang, T.H., Thanh, N.T.N., Luyen, B.T.T., Huong, T.T., Wilson, J., Beutler, J.A., Ban, N.K., Cuong, N.M. and Kim, Y.H. (2011) An Evaluation of the RNase H Inhibitory Effects of Vietnamese Medicinal Plant Extracts and Natural Compounds. *Pharmaceutical Biology*, **49**, 1046-1051. <http://dx.doi.org/10.3109/13880209.2011.563316>
- [11] Kumar, S., Kumar, D., Deshmukh, R.R., Lokhande, P.D., More, S.N. and Rangari, V.D. (2008) Antidiabetic Potential of *Phyllanthus reticulatus* in Alloxan-Induced Diabetic Mice. *Fitoterapia*, **79**, 21-23. <http://dx.doi.org/10.1016/j.fitote.2007.06.012>
- [12] Kumar, S., Kumar, D., Deshmukh, R.R. and Rangari, V.D. (2007) Hypoglycemic Activity of Roots of *Phyllanthus reticulatus* in Alloxan Induced Diabetic Mice. *International Journal of Plant Science*, **2**, 184-187.
- [13] Hui, W.H., Li, M.M. and Wong, K.M. (1976) A New Compound, 21 α -Hydroxyfriedel-4(23)-en-3-One and Other Triterpenoids from *Phyllanthus reticulatus*. *Phytochemistry*, **15**, 797-798. [http://dx.doi.org/10.1016/S0031-9422\(00\)94448-0](http://dx.doi.org/10.1016/S0031-9422(00)94448-0)
- [14] Lan, M.S., Ma, J.X., Tan, C.H., Wei, S. and Zhu, D.Y. (2010) Chemical Constituents of *Phyllanthus reticulatus*. *Helvetica Chimica Acta*, **93**, 2276-2280. <http://dx.doi.org/10.1002/hlca.201000168>
- [15] Pojchaijongdee, N., Sotanaphun, U., Limsirichaikul, S. and Poobrasert, O. (2010) Geraniinic Acid Derivative from the Leaves of *Phyllanthus reticulatus*. *Pharmaceutical Biology*, **48**, 740-744. <http://dx.doi.org/10.3109/13880200903273898>
- [16] Ma, J.X., Lan, M.S., Qu, S.J., Tan, J.J., Luo, H.F., Tan, C.H. and Zhu, D.Y. (2012) Arylnaphthalene Lignan Glycosides and Other Constituents from *Phyllanthus reticulatus*. *Journal of Asian Natural Products Research*, **14**, 1073-1077. <http://dx.doi.org/10.1080/10286020.2012.712040>
- [17] Begum, T., Rahman, M.S. and Rashid, M.A. (2006) Phytochemical and Biological Investigations of *Phyllanthus reticulatus*. *Dhaka University Journal of Pharmaceutical Sciences*, **5**, 21-23.
- [18] Lam, S.H., Wang, C.Y., Chen, C.K. and Lee, S.S. (2007) Chemical Investigation of *Phyllanthus reticulatus* by HPLC-SPE-NMR and Conventional Methods. *Phytochemical Analysis*, **18**, 251-255. <http://dx.doi.org/10.1002/pca.979>
- [19] Sharma, S. and Kumar, S. (2013) *Phyllanthus reticulatus* Poir.—An Important Medicinal Plant: A Review of Its Phytochemistry, Traditional Uses and Pharmacological Properties. *International Journal of Pharmaceutical Sciences & Research*, **4**, 2528-2534.
- [20] Jamal, A.K., Yaacob, W.A. and Din, L.B. (2008) A Chemical Study on *Phyllanthus reticulatus*. *Journal of Physical Science*, **19**, 45-50.
- [21] Ogundajo, A.L., Ogunwande, I.A., Bolarinwa, T.M., Joseph, O.R. and Flamini, G. (2014) Essential Oil from the

- Leaves of *Hibisciscus suranthesis* L. from Nigeria. *Journal of Essential Oil Research*, **26**, 114-117. <http://dx.doi.org/10.1080/10412905.2013.860410>
- [22] British Pharmacopoeia (1980) Vol II, H.M. Stationary Office.
- [23] Adams, R.P. (2007) Identification of Essential Oil Components by Gas Chromatography/Quadrupole Mass Spectrometry. 4th Edition, Allured Publishing Corporation, Carol Stream.
- [24] Joulain, D. and Koenig, W.A. (1989) The Atlas of Spectral Data of Sesquiterpene Hydrocarbons. E.B. Verlag, Hamburg.
- [25] National Institute of Standards and Technology (2011) Chemistry Web Book. Data from NIST Standard Reference Database 69. <http://www.nist.gov/>
- [26] Villarreal, S., Bojas, L.B., Usubillaga, A., Ramirez, I. and Solorzano, M. (2008) Volatile Constituents from the Leaves of *Phyllanthus salviaefolius* H.B.K. from the Venezuelan Andes. *Natural Product Communications*, **3**, 275-277.
- [27] Moronkola, D.O., Ogunwande, I.A., Oyewole, I.O., Başer, K.H.C., Ozek, T. and Ozek, G. (2009) Studies on the Volatile Oils of *Momordica charantia* L. (Cucurbitaceae) and *Phyllanthus amarus* Sch. & Thonn. (Euphorbiaceae). *Journal of Essential Oil Research*, **21**, 393-399. <http://dx.doi.org/10.1080/10412905.2009.9700201>
- [28] Quijano, C.E., Linares, D. and Pino, J.A. (2007) Changes in Volatile Compounds of Fermented Cereza Agria [*Phyllanthus acidus* (L.) Skeels] Fruit. *Flavour and Fragrance Journal*, **22**, 392-394. <http://dx.doi.org/10.1002/ffj.1810>
- [29] Li, X., Wei, W.X. and Lin, C.W. (2008) Comparative Analysis of Essential Oil Compositions from *Phyllanthus niruru*, *P. urinaria*, and *P. arenarius*. *Chemistry of Natural Compounds*, **44**, 257-259.
- [30] Liu, X.L., Zhao, M.M., Luo, W., Yang, B. and Jiang, Y.M. (2009) Identification of Volatile Components in *Phyllanthus emblica* L. and Their Antimicrobial Activity. *Journal of Medicinal Food*, **12**, 423-428. <http://dx.doi.org/10.1089/jmf.2007.0679>
- [31] Liu, X.L., Zhao, M.M., Wang, J.S. and Wei, L. (2008) Effectiveness of *Phyllanthus emblica* L. Essential Oil to Inhibit the Growth of Food-Spoiling Yeasts. *Journal of Food Safety*, **28**, 261-275. <http://dx.doi.org/10.1111/j.1745-4565.2008.00119.x>
- [32] Brusotti, G., Cesari, I., Gilardoni, G., Tosi, S., Grisoli, P., Picco, A.M. and Caccialanza, G. (2012) Chemical Composition and Antimicrobial Activity of *Phyllanthus muellerianus* (Kuntze) Excel Essential Oil. *Journal of Ethnopharmacology*, **142**, 657-662. <http://dx.doi.org/10.1016/j.jep.2012.05.032>

Scientific Research Publishing (SCIRP) is one of the largest Open Access journal publishers. It is currently publishing more than 200 open access, online, peer-reviewed journals covering a wide range of academic disciplines. SCIRP serves the worldwide academic communities and contributes to the progress and application of science with its publication.

Other selected journals from SCIRP are listed as below. Submit your manuscript to us via either submit@scirp.org or [Online Submission Portal](#).

