

Genetic Diversity and Population Structure of North China Mountain Walnut Revealed by ISSR

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Abstract

North China Mountain Walnut (NCMW) is one of the ancestors of extant cultivated species, and a valuable gene resource for resistance breeding of walnut in China. Inter-Simple Sequence Repeat (ISSR) primers were designed to evaluate the level and pattern of genetic diversity in eight populations of NCMW. Nine ISSR primers yielded 91 amplification products with different sizes, of which 84 (92.31%) were polymorphic. A high species-level genetic diversity was detected with Nei's ($H = 0.2592$) and Shannon's diversity ($I = 0.4003$). In contrast, the population-level genetic diversity was relatively lower (PPB = 43.27%, $H = 0.1347$, $I = 0.1862$). Coefficient of populations differentiation (G_{ST}) was 0.5066, indicating that inter-population and intra-population variation contributed 50.66% and 49.34% respectively to the total genetic variability. This relative level of variation was further supported by AMOVA analysis. Limited gene flow ($Nm = 0.5133$), habitat fragmentation and geographical isolation might be responsible for the population structure of NCMW. UPGMA cluster analysis classified the eight populations into three groups which showed no significant relationship between the genetic similarity coefficient and geographic origin but showed remarkable association with morpho-physiological characters, particularly nut traits. The results of the study provide species-level and population-level genetic profiles for further exploitation and conservation of genetic diversity of NCMW.

Keywords

North China Mountains Walnut (NCMW), Genetic Diversity, Population Structure, ISSR

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1. Introduction

Several walnut (*Juglans spp.*) species are important dual purpose fruit and timber economic forest tree species, of these the globally most important is “Common Walnut” (*J. regia*). China is the one of origin centre of walnuts and possesses abundant resources. There are five main species of the *Juglans* genus available as walnut breeding resources in China. These are *J. regia* L., *J. Sigillata* Dode, *J. mandshurica* Maxim, *J. openensis* Hu, and *J. nigra* L. [1]. Due to walnut being seed propagated with open-pollination, walnut is highly variable with a diverse gene pool [2]. Walnuts in China have been divided into four ecotypes (Sinkiang Walnut, North China Mountain Walnut, Qinling-Daba Mountain Walnut and the Tibetan Highlands Walnut) in terms of their geographical distribution [3]. Genetic diversity research on North China Mountain Walnuts has developed slowly in comparison with the other three ecotypes in China.

In China, North China Mountain Walnuts are mainly distributed in the foothills of the North China Mountains, which spread across the northern latitudes from 35° to 40°, and the western longitudes from 112° to 119°. The area is a rocky, mountainous region with serious soil erosion, a warm temperate climate, an annual precipitation from 400 - 700 mm, and slightly calcareous cinnamon soils [1]. Owing to the unique geography and climate, the endemic walnut varieties contain many agronomically excellent properties and specific valuable genes, such as for high content of protein, and strong disease and drought resistances [3], which have significant potential value for walnut variety improvement. In addition, the North China Mountains could be where *J. regia* L. and *J. mandshurica* Maxim were first domesticated and the region is one of the cultivated species' provenance [3]. Thus, the North China Mountain Walnut could possess a vast gene resource. However, in the past decade, under long-term biological or environmental pressure, the native walnut resources have been seriously damaged, which may result in increased homogeneity or reduction of genetic variability. The lack of systematic studies of genetic diversity among Chinese *Juglans* species and ecotypes could seriously restrict genetic improvement by limiting exploitation in walnut culture and breeding of many excellent traits found in these landrace lines. Accordingly, it is essential to properly characterize and assess the genetic diversity of landrace walnut resources for protection and breeding utilization.

In the past few years, a quantity of molecular markers, such as RFLP (Restriction Fragment-Length Polymorphism) [4] [5], RAPD (Randomly Amplified Polymorphic DNA) [6]-[9], AFLP (Amplified Fragment Length Polymorphism) [10] [11], SSR (Simple Sequence Repeat or microsatellites) [9] [12]-[17], and ISSR (Intersimple Sequence Repeat) [18] have been generally employed to examine genetic diversity and structure in walnut. While previous reports concentrated on the fingerprinting and genetic variability of walnut germplasm, there are few studies on the population genetic diversity of different ecotypes [16] [19] [20], especially the North China Mountain Walnut. ISSRs have been widely employed to evaluate genetic diversity of populations of economic forest tree species [21]-[23].

It is reported for the first time to assess genetic diversity and genetic structure of North China Mountain Walnut sampled out of eight significant natural populations in China by ISSR. We evaluate the population-level and species-level genetic diversity of eight North China Mountain Walnut populations throughout their known distribution by ISSR analysis to give some references for conservation measures and germplasm resource development and utilization.

2. Materials and Methods

2.1. Plant Material and Sampling Strategy

In this study, a total of 138 individuals of North China Mountain Walnut, representing eight populations (GX, FY, LY, YX, LC, SX, ZQ, and LZ), were sampled across three provinces in China, including Shanxi, Hebei and Henan along the Taihang and Lvliang Mountains (Figure 1 and Table 1). The collection locations represented a wide geographic distribution of North China Mountain Walnut. Interpopulation distance ranged from a minimum of 30.8 km (LC-SX) to a maximum of 512 km (LZ-LY). The number of samples analyzed depended on the usability of adult plants (>80 cm at breast height diameter) in each sampling location. In each population, 4 - 25 adult walnut trees were randomly sampled at the distance exceeding at least 50 m from each other to reduce the probability of sampling from family clusters. From each individual tree, fresh mature leaves were randomly sampled, stored in plastic bags and taken to the lab for experimental studies.

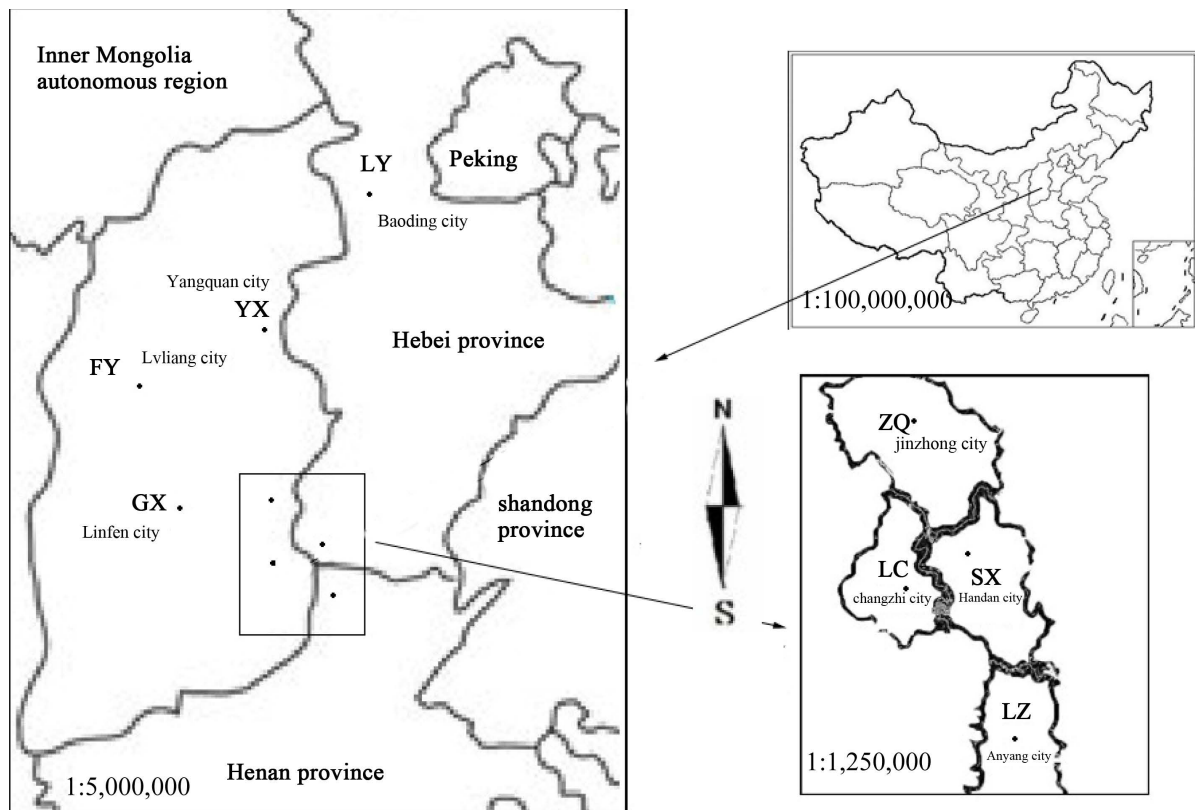


Figure 1. Geographic locations of sampling sites for the eight natural populations of North China Mountain Walnut examined in this study. The abbreviations of GX, FY, LY, YX, LC, SX, ZQ and LZ follow those used in **Table 1**.

Table 1. Location and sampling size of eight nature populations of North China Mountains walnut.

Population	Location	Longitude (E)/Latitude (N)	Elevation (m)	Sample sizes
GX	Gu County, Linfen City, Shanxi Province	36°32'N 112°04'E	1430	15
FY	Fengyang County, Luliang City, Shanxi Province	37°15'N 111°37'E	1047	15
ZQ	Zunquan County, Jinzhong City, Shanxi Province	36°51'N 113°27'E	821	23
YX	Yu County, Yangquan City, Shanxi Province	38°23'N 113°21'E	653	20
LC	Licheng County, Changzhi City, Shanxi Province	36°48'N 113°27'E	757	25
SX	Shexian County, Handan City, Hebei Province	36°41'N 113°27'E	562	11
LY	Laiyuan County, Baoding City, Hebei Province	39°53'N 118°53'E	45	8
HN	Linzhou County, Anyang City, Henna Province	36°16'N 113°48'E	420	4

2.2. DNA Extraction

Genomic DNA was extracted from storage leaves of walnut by using the optimized Cetyl Trimethyl Ammonium Bromide (CTAB) method [11]. Total DNA quality was checked by gel electrophoresis and DNA concentration was determined by Biophotometer protein nucleotide analyzer (Eppendorf, Germany). Purified DNA was diluted to approximately $30 \text{ ng}\cdot\mu\text{L}^{-1}$ prior to polymerase chain reaction (PCR) amplification, and preserved in the refrigerator at -20°C .

2.3. ISSR-PCR Amplification

A total of one hundred ISSR primers were used. The primers were designed according to public sequences pro-

vided by the University of British Columbia, Canada and biosynthesised by the Shanghai Sangon Biological Engineering Technology & Services Co. Ltd. (Shanghai, China). They were sifted firstly grounded on one sample from each population. Nine primers, which yielded clear, polymorphic, and reproducible bands, were used for the further genetic diversity analysis of all 138 DNA individuals (**Table 2**). PCR reactions were performed in 25 μ l reaction mixture containing 2.5 μ l of 10 \times buffer, 1 U *Taq* polymerase, 0.2 μ M dNTP (Tiangen Biotech (Beijing) Co., Ltd), 0.2 μ M primer and 30 ng of genomic DNA. Amplification was carried out in a thermocycler (Peltier Thermal Cycler PTC-200, MJ research, USA) as follows: initial denaturation of 5 min at 94°C, followed by 35 cycles of denaturation of 30 s at 94°C, annealing of 45 s at 51°C - 60°C (based on the used primer), and extension of 1.5 m at 72°C, followed by a final extension of 7 m at 72°C. The amplification products were separated on 2.0% agarose gels in 0.5 \times TBE buffer at a constant voltage of 80 V, for 2 h along with 100 bp DNA ladder marker (100 - 1500 bp) and stained by 0.1% ethidium bromide. The gel images were then visualized and recorded using a Bio-Rad Gel Doc 2000TM system (USA).

2.4. Data Analysis

Polymorphic bands were scored visually as either present “1” or absent “0” to form a binary matrix. On the supposition of Hardy-Weinberg equilibrium, POPGENE versions 1.31 [24] were employed to compute the following basic statistics. To assess genetic diversity at species level and population level, the parameters calculated included; percentages of polymorphic bands (*PPB*%), mean number of alleles (*Na*), mean effective number of alleles (*Ne*), Nei’s gene diversity index (*H*) and Shannon-diversity index (*I*). To evaluate gene differentiation between populations, the parameters calculated included Nei’s coefficient of population differentiation (*G_{ST}*) with the formula: $G_{ST} = (Ht - Hs)/Ht$ [25], and Gene flow among populations (*Nm*) which was calculated using the formula of $Nm = (1 - G_{ST})/2G_{ST}$. Genetic relationships among populations were estimated by Nei’s unbiased genetic distances (*D*) and genetic identities (*I*) using NTSYSpc-2.0 [26], and a cluster analysis was performed using UPGMA (the Unweighted Pair Group Method with Arithmetic mean) [27]. In addition, population differentiation coefficients within and among the populations were computed from AMOVA 1.55 (analysis of molecular variance) [28].

3. Results

3.1. Genetic Diversity

91 bands in total were generated from nine screened primers for the 138 individual samples from the eight populations, of which 84 (92.13%) were polymorphic bands. The size of the amplified band ranged from 300 bp to 1200 bp. The percentage of polymorphic band (*PPB*) of the 9 ISSR primers differed from 66.67% (UBC855) to 100% (UBC815, UBC836, UBC840 and UBC853), with an average of 92.31%. The number of bands presented by each primer also differed from 6 (UBC855) to 11 (UBC840) with an average of 10.11 (**Table 2**).

Estimates of genetic diversity in eight populations of North China Mountain Walnut are summarized in **Table 3**.

Table 2. Polymorphic bands generated by ISSR in eight nature populations of NCMW.

Primer	Sequence ^a	Tmb/ ^o C	Number of band	Number of polymorphic band	PPB/%
UBC811	(GA)8C	54.7	11	9	81.82
UBC815	(CT)8A	59.1	10	10	100
UBC836	(AG)8YA	52.5	8	8	100
UBC840	(GA)8YT	50.6	15	15	100
UBC843	(CT)8RC	54.3	10	8	80
UBC853	(TC)8RT	55.2	11	11	100
UBC855	(AC)8YT	51.8	6	4	66.67
UBC856	(AC)8YA	56.1	9	9	100
UBC888	BDB(CA)7	53.4	11	10	90.91
	Average		10.11	9.33	92.31

Y = (C,T), R = (A,G), B = (C,G,T), D = (A,G,T). ^aSequence 5' - 3'; ^bPCR reaction annealing temperature for tabulated primers (^oC); ^cPercentage of polymorphic band.

Table 3. Genetic diversity analysis of 8 populations of North China Mountain based on ISSR markers.

Population	Number of polymorphic loci	PPB (%)	N_a	N_e	H	I
GX	48	52.75	1.575	1.3024	0.1788	0.2695
YX	30	32.97	1.3297	1.1505	0.0955	0.1496
LY	38	41.76	1.4176	1.1795	0.1098	0.1716
FY	52	57.14	1.5714	1.3163	0.1839	0.2778
LC	39	42.86	1.4286	1.1769	0.1090	0.1717
SX	41	45.05	1.4505	1.2764	0.1617	0.2413
ZQ	49	53.85	1.5385	1.2584	0.1569	0.2433
HN	18	19.78	1.1978	1.1419	0.0818	0.1192
Average	39.375	43.27	1.4386	1.2253	0.1347	0.1862
Species	84	92.31	1.9231	1.4243	0.2592	0.4003
SD			0.2679	0.3335	0.1699	0.2290

PPB, Percentage of polymorphic loci (%); N_a , Observed number of alleles; N_e , Effective number of alleles; H , Nei's genetic diversity; I , Shannon's information index; SD , Indicates the standard deviation.

At the species level, the effective number of alleles (N_e), Nei's genetic diversity (H) and the Shannon information index (I) were 1.4243, 0.2592, 0.4003 respectively. At the population level, PPB ranged from 19.78% for the LZ population to 57.14% for the YX population, with an average of 43.27%, and the average N_e , H , and I were 1.2253, 0.1347, and 0.1862, respectively. The results showed that the genetic diversity of North China Mountain Walnuts from the YX population ($N_e = 1.3163$, $H = 0.1839$, $I = 0.2778$) was the richest in three estimates among the eight populations and the lowest diversity was exhibited in the LZ population.

3.2. Genetic Differentiation

Nei's coefficient of genetic differentiation (G_{ST}) was 0.5066, which showed that 50.66% of the total genetic variability is attributed to inter-populations and 49.34% intra-populations. Results of AMOVA also agreed with which genetic differentiation inter-populations is relatively high ($F_{ST} = 0.5161$). At two hierarchical levels, 51.61% of the total genetic variation was partitioned inter-populations, and 48.39% ($P < 0.001$) intra-populations. The level of gene flow (Nm) was estimated to be only 0.5133, which implied that a low gene flow would not prevent genetic drift thus enabling the gene differentiation between populations.

3.3. Genetic Similarity and Genetic Distance among Populations

To further clarify the gene differentiation among different populations, Nei's pairwise genetic similarity coefficients were assessed. Genetic similarity coefficients varied from 0.7423 to 0.9298 with an average 0.8159, which showed higher levels of genetic differentiation partitioned among populations of North China Mountain Walnut. The highest similarity coefficient was between the ZQ population and the LC population, while difference in similarity coefficients between the FY population and the LC population was revealed as the lowest (Table 4).

In order to further illustrate relationships among populations, a dendrogram generated by UPGMA algorithm based on Nei's genetic similarity, which clustered the eight populations into three major groups (Figure 2). The first group included five populations (ZQ, LC, SX, YX and LY), distributed along the main vein of Taihang Mountains, the second group comprised GX and FY populations, located in the south of Taiyue Mountains and east of Lvliang Mountains respectively, while LZ alone formed the third group, located in the east of Taihang Mountains. However, the cluster result was not necessarily relationship with geographic separation. Such as, the ZQ population had a much greater geographic separation from the LY population than the LZ population, but ZQ clustered together with the LY population and not with the LZ population in the dendrogram. Therefore, geographical distance alone cannot account for the genetic distance between populations.

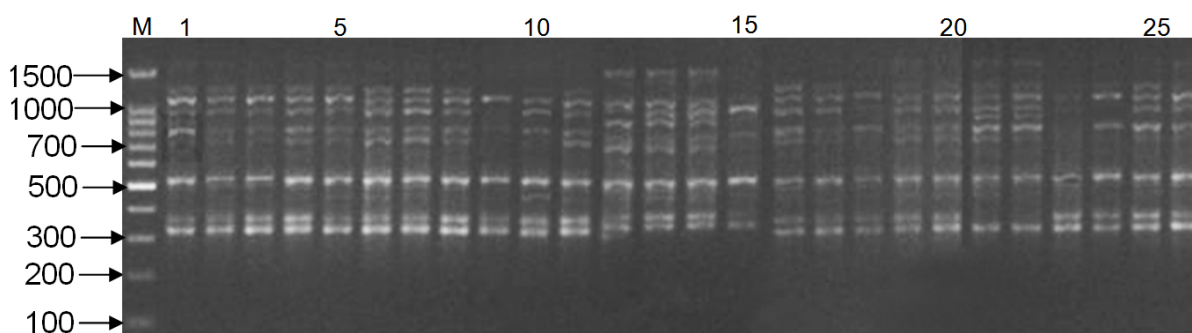
4. Discussion

4.1. Genetic Diversity

Our data shows that genetic diversity is higher at the species level ($PPB = 92.31\%$, $H = 0.2592$, $I = 0.4003$), but

Table 4. Nei's unbiased genetic identity (above diagonal) and genetic distance (below diagonal) among eight nature populations of North China walnut.

Population	GX	FY	LY	YX	LC	SX	ZQ	HN
GX	****	0.8567	0.7954	0.7892	0.8077	0.8337	0.7913	0.8024
FY	0.1546	****	0.7658	0.7451	0.7423	0.7562	0.7653	0.7562
LY	0.2289	0.2668	****	0.8751	0.8891	0.8735	0.8740	0.8424
YX	0.2367	0.2943	0.1334	****	0.9011	0.8868	0.8709	0.8350
LC	0.2136	0.2979	0.1176	0.1041	****	0.9102	0.9298	0.8259
SX	0.1818	0.2794	0.1353	0.1202	0.0941	****	0.8993	0.8454
ZQ	0.2340	0.2675	0.1346	0.1382	0.0728	0.1062	****	0.8761
HN	0.2201	0.2795	0.1715	0.1804	0.1913	0.1679	0.1322	****

**Figure 2.** ISSR amplification results with primer UBC811 for GX population.

lower at population level ($PPB = 43.27\%$, $H = 0.1347$, $I = 0.1862$), which is alike to the results obtained from same species using different molecular markers [17] [19] [20]. When compared with many other economic tree species using ISSR, North China Mountain Walnut maintains higher species diversity (e.g. ginkgo [29], *Emmenopterys henryi* [30], etc.). The higher species diversity may largely be attributed to walnut breeding systems. The walnut is wind pollinated. When compared with insect pollination, the smaller wind carried pollen is easier to spread. In addition, walnut has unisexual flowers and a monoecious plant, which may carry out both geitonogamy and cross pollination, but due to dichogamy, the same cultivars or strains while self fertile may not be able to pollinate itself, which results in frequent gene flow among individuals and increases the chance of gene recombination. Geographical distribution would be another key factor that determines genetic diversity level of a species. The North China Mountain Walnut is distributed mostly in rocky Mountainous areas, which have serious soil erosion and calcareous cinnamon soils. The need and hence ability to adapt to different environments results in a higher level of genetic diversity.

North China Mountain Walnuts are distributed in different ecological environments and geographic conditions, such as having a range of growing elevations from 45 m to 1430 m above sea level. With increasing elevation, climate and soil physicochemical properties also show a significant change, which results in greater population variation. As observed in our study, the population differentiation parameters such as PPB , Ne , H and I in eight populations showed a similar pattern, the YX population is the highest, and the LZ population is the lowest. This can be interpreted as a consequence of habitat fragmentation and small numbers of individuals. Due to genetic drift, small populations tend to lose genetic variation has been verified in many studies. Therefore, the results need to be further studied and verified by a more extensive investigation.

Compared with other ecotypes, genetic diversity of North China Mountain Walnut was at a moderate level, which was marginally higher than Sinkiang walnut and Qinling-Daba Mountain walnut [31], but was lower than Tibet walnut [17]. The result of this study supported the conclusion that the Tibet walnut population diversity was generally higher than other ecotypes populations [17].

4.2. Populations Genetic Structure

In accordance with the G_{ST} value ($G_{ST} = 0.5066$), found in this study a significant amount of genetic differentia-

tion is observed among and within eight populations of North China Mountain Walnut. This point is reinforced by the AMOVA, which indicating that 51.61% of the genetic variation was partitioned inter-populations and 49.34% intra-populations. Significant genetic differentiation was also reported by Wu *et al.* using RAPD. In contrast, less genetic differentiation was found by Xi [3] using a bioecological survey method and peroxidase isoenzymatics marker. The difference might be due to the sampling locations, sampling size, and the different marker systems used. Similar studies have shown that there are inconsistencies in genetic differentiation value found between populations when using different genetic markers in other species populations [32] [33].

High genetic differentiation of North China Mountain Walnut is attributed to the following points: Firstly, because of low gene flow of the North China Mountain Walnut habitat ($Nm = 0.5133$), the present population structure was shaped by genetic drift mainly. Secondly, the Taihang and Lvliang Mountains contribute to the geographical isolation, which severely hindered dispersal of pollen or seeds and exchange of genes between populations, ultimately promoting the large genetic differentiation among populations. Thirdly, in order to adapt to the diverse habitats of the region in the evolutionary process, variants to some extent occurred, which may have been preserved and fixed gradually due to limited gene flow, thereby, genetic differentiation among populations occurred. The G_{ST} value and AMOVA analysis results obtained in this study indicated that North China Mountain Walnut has a significant genetic differentiation among populations, which suggests that the species has a strong environmental adaptability. Put briefly, high genetic variation among populations is due to genetic drift and geographical isolation of the populations.

Based on the genetic similarity, eight populations of North China Mountain Walnut were grouped into three clusters, but genetic differentiation did not completely cluster according to geographical distances (Figure 3). This supported the conclusion that owing to the complex origin and affiliation of walnut, there isn't an obvious association between genetic similarity and geographic origin [11]. But the results of cluster analysis did correspond with phenetic classification systems of local varieties. Mian walnut, with a less developed endoseptum, a slightly colored and a slightly veiny shell, is the main variety in the LC, ZQ and SX populations; it is characterized by large fruit size, thin and smooth shell and pale yellow kernels in the LY and YX population; the Fenzhou and Guxian walnut is well known to have a thin shell, full kernels, higher kernel crackout, milky white kernels in the FY and GX population, respectively; the varieties of the LZ population are Mian walnut and Jia walnut with thicker, corrugated and tawny shell and low kernel crackout. Whether these differences are due to genetic drift still needs to be determined.

4.3. Conservation Implications

To know about the geographic structure of intraspecific genetic variation is indispensable for the protection of germplasm resources. Our study provides an insight into genetic diversity and genetic differentiation at population levels of North China Mountain Walnut, and indicates a decrease of genetic diversity caused by genetic drift is unacceptable. Accordingly, so as to secure existing diversity, preservation areas covering large popula-

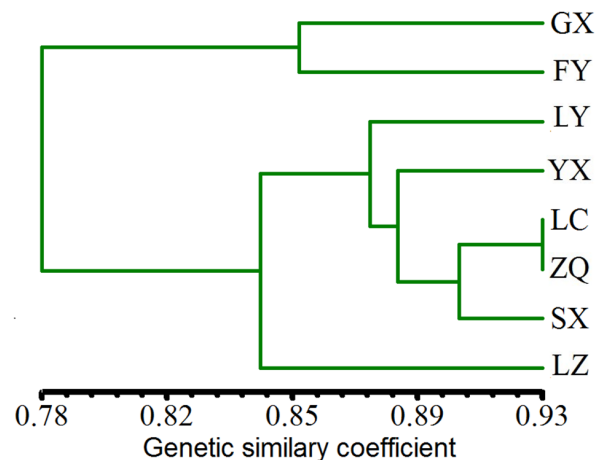


Figure 3. UPGMA dendrogram for 8 populations of North China Mountain Walnut based on genetic identity.

tions as well as many small populations should be established. Mixing genetically diverse populations of North China Mountain Walnut in situ or ex situ is unacceptable in conservation.

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References

- [1] Wu, G.L., Liu, Q.L., Zheng, X.B., Song, Y.Q., Jian, Z.H. and Peng, G.B. (2009) Advances in Research on the Worldwide Walnut Germplasm. *Journal of Fruit Science*, **26**, 539-545. (In Chinese)
- [2] Ebrahimi, A., Fatahi, R. and Zamani, Z. (2011) Analysis of Genetic Diversity among Some *Persian walnut* Genotypes (*Juglans regia* L.) Using Morphological Traits and SSRs Markers. *Scientia Horticulturae*, **130**, 146-151. <http://dx.doi.org/10.1016/j.scienta.2011.06.028>
- [3] Xi, S.K. (1987) Gene Resources of *Juglans* and Genetic Improvement of *Juglans regia* in China. *Scientia Silvae Sinicae*, **23**, 342-349. (In Chinese)
- [4] Fjellstrom, R.G., Parfitt, D.E. and McGranahan, G.H. (1994) Genetic Relationship and Characterization of *Persian walnut* (*Juglans regia* L.) Cultivars Using Restriction Fragment Length Polymorphisms (RFLPs). *Journal of the American Society for Horticultural Science*, **119**, 833-839.
- [5] Fjellstrom, R.G. and Parfitt, D.E. (1994) Walnut (*Juglans* spp.) Genetic Diversity Determined by Restriction Fragment Length Polymorphisms. *Genome*, **37**, 690-700. <http://dx.doi.org/10.1139/g94-097>
- [6] Nicese, F.P., Hormaza, J.I. and McGranahan, G.H. (1998) Molecular Characterization and Genetic Relatedness among Walnut (*Juglans regia* L.) Genotypes Based on ISSR Markers. *Euphytica*, **101**, 199-206. <http://dx.doi.org/10.1023/A:1018390120142>
- [7] Wu, Y.M., Pei, D., Xi, S.K. and Li, J.R. (2000) A Study on the Genetic Relationship among Species in *Juglans* L. Using RAPD Markers. *Acta Horticulturae Sinica*, **27**, 17-22. (In Chinese)
- [8] Wu, Y.M., Liu, Y., Dong, F.X. and Xi, S.K. (2000) Study on Different Ecological Types of Chinese Walnut (*J. regia*) Using RAPD Markers. *Journal of Beijing Forestry University*, **22**, 23-27. (In Chinese)
- [9] Nazeer, A., Mir, J.I., Reyazul, R.Mir., Nazir, A.R., Rizwan, R., Shabir, H.W., Wajida, S., Hidayatullah, M. and Sheikh, M.A. (2012) SSR and RAPD Analysis of Genetic Diversity in Walnut (*Juglans regia* L.) Genotypes from Jammu and Kashmir, India. *Physiology and Molecular Biology of Plants*, **18**, 149-160. <http://dx.doi.org/10.1007/s12298-012-0104-z>
- [10] Bayazit, S., Kazan, K., Gulbitti, V., Evik, C., Ayanoglu, H. and Ergul, A. (2007) AFLP Analysis of Genetic Diversity in Low Chilling Requiring Walnut (*Juglans regia* L.) Genotypes from Hatay, Turkey. *Scientia Horticulturae*, **111**, 394-398. <http://dx.doi.org/10.1016/j.scienta.2006.11.006>
- [11] Wang, H.X., Zhao, S.G., Gao, Y., Zhang, Z.H. and Xuan, L.C. (2011) Genetic Diversity of *Juglans regia* L. Cultivars Revealed by AFLP Analysis. *Scientia Agricultura Sinica*, **44**, 1434-1442. (In Chinese)
- [12] Dangl, G.S., Woeste, K., Aradhya, M.K., Koehmstedt, A., Simon, C., Potter, D., Leslie, C.A. and McGranahan, G. (2005) Characterization of 14 Microsatellite Markers for Genetic Analysis and Cultivar Identification of Walnut. *Journal of the American Society for Horticultural Science*, **130**, 348-354.
- [13] Foroni, I., Rao, R., Woeste, K. and Gallitelli, M. (2005) Characterization of (*Juglans regia* L.) with SSR Markers and Evaluation of Genetic Relationships among Cultivars and the "Sorrento" Landrace. *The Journal of Horticultural Science and Biotechnology*, **80**, 49-53.
- [14] Victory, E.R., Jeffrey, C., Glaubitz, O.E., Rhodes, J.R. and Woeste, K.E. (2006) Genetic Homogeneity in *Juglans nigra* (Juglandaceae) at Nuclear Microsatellites. *American Journal of Botany*, **93**, 118-126. <http://dx.doi.org/10.3732/ajb.93.1.118>
- [15] Wang, H., Hao, J.M., Wang, B.Q. and Pei, D. (2007) SSR Analysis of Genetic Diversity of Eight Natural Walnut Populations in China. *Scientia Silvae Sinicae*, **43**, 120-124. (In Chinese)
- [16] Wang, H., Pei, D., Gu, R.S. and Wang, B.Q. (2008) Genetic Diversity and Structure of Walnut Populations in Central and Southwestern China Revealed by Microsatellite Markers. *Journal of the American Society for Horticultural Science*, **1332**, 197-203.

- [17] Wang, H. (2010) Genetic Diversity of Germplasm Resources on Walnut in Tibet Region. Thesis, Chinese Academy of Forestry Sciences, Peking, 77-78. (In Chinese)
- [18] Potter, D., Gao, F., Aiello, G., Leslie, C. and McGranahan, G. (2002) Intersimple Sequence Repeat Markers for Fingerprinting and Determining Genetic Relationships of Walnut (*Juglans regia*) Cultivars. *Journal of the American Society for Horticultural Science*, **127**, 75-81.
- [19] Christopoulou, M.V., Dimos, R., Eleni, T. and Penelope, J.B. (2010) Germplasm Diversity and Genetic Relationships among Walnut (*Juglans regia* L.) Cultivars and Greek Local Selections Revealed by Inter-Simple Sequence Repeat (ISSR) Markers. *Scientia Horticulturae*, **125**, 584-592. <http://dx.doi.org/10.1016/j.scienta.2010.05.006>
- [20] Li, C., Luo, S.P., Zeng, B., Li, J. and Li, G. (2011) Analysis of Genetic Diversity of Germplasm Resources of Walnut (*Juglans regia* L.) Revealed by ISSR in Xinjiang of China. *Scientia Agricultura Sinica*, **44**, 1871-1879.
- [21] Thimmappaiah, W.G., Santhosh, D., Shobha, G.S. and Melwyn, G.S. (2009) Assessment of Genetic Diversity in Cashew Germplasm Using RAPD and ISSR Markers. *Scientia Horticulturae*, **120**, 411-417. <http://dx.doi.org/10.1016/j.scienta.2008.11.022>
- [22] Ai, C.X., Zhang, L.S., Wei, H.R., Jin, S.N., Yuan, K.J. and Liu, Q.Z. (2007) Study on the Genetic Diversity of Natural Chestnut of Shandong by ISSR. *Chinese Journal of Biotechnology*, **23**, 628-633. (In Chinese) [http://dx.doi.org/10.1016/S1872-2075\(07\)60043-0](http://dx.doi.org/10.1016/S1872-2075(07)60043-0)
- [23] Kamallesh, S.M., Tikam, S.R. and Shirish, A.R. (2011) Molecular Analyses of Genetic Variability in Soap Nut (*Sapindus mukorossi* Gaertn.). *Industrial Crops and Products*, **34**, 1111-1118. <http://dx.doi.org/10.1016/j.indcrop.2011.03.029>
- [24] Yeh, F.C., Yang, R.C. and Boyle, T. (1999) POPGENE Version 1.32: Microsoft Window-Based Freeware for Population Genetics Analysis. University of Alberta, Edmonton.
- [25] Nei, M. (1973) Analysis of Gene Diversity in Subdivided Populations. *Proceedings of the National Academy of Sciences of the United States of America*, **70**, 3321-3323. <http://dx.doi.org/10.1073/pnas.70.12.3321>
- [26] Rohlf, F.J. (2000) NTSYS-Pc: Numerical Taxonomy and Multivariate Analysis System. Version 2.1., Exeter Publishing Ltd., New York.
- [27] Nei, M. (1978) Estimation of Average Heterozygosity and Genetic Distance from a Small Number of Individuals. *Genetics*, **89**, 583-590.
- [28] Excoffier, L., Smouse, P. and Quattro, J.M. (1992) Analysis of Molecular Variance Inferred from Metric Distances among DNA Haplotypes: Application to Human Mitochondrial DNA Restriction Data. *Genetics*, **131**, 479-491.
- [29] Ge, Y.Q., Qiu, Y.X., Ding, B.Y. and Fu, C.X. (2003) An ISSR Analysis on Population Genetic Diversity of the Relict Plant *Ginkgo biloba*. *Biodiversity Science*, **11**, 276-287. (In Chinese)
- [30] Li, J.M. and Jin, Z.C. (2008) Genetic Structure of Endangered *Emmenopterys henryi* Oliv. Based on ISSR Polymorphism and Implications for Its Conservation. *Genetica*, **133**, 227-234. <http://dx.doi.org/10.1007/s10709-007-9204-z>
- [31] Li, G.T., Ai, C.X., Zhang, L.S., Wei, H.R. and Liu, Q.Z. (2011) ISSR Analysis of Genetic Diversity among Seedling Walnut (*Juglans* spp.) Populations. *Journal of Plant Genetic Resources*, **12**, 640-645. (In Chinese)
- [32] Freville, H., Justy, F. and Olivieri, I. (2001) Comparative Allozyme and Microsatellite Population Structure in a Narrow Endemic Plant Species, *Centaurea corymbosa* Pourret (Asteraceae). *Molecular Ecology*, **10**, 879-889. <http://dx.doi.org/10.1046/j.1365-294X.2001.01249.x>
- [33] Maguire, T.L., Peakall, R. and Saenger, P. (2002) Comparative Analysis of Genetic Diversity in the Mangrove Species *Avicennia marina* (Forsk.) Vierh. (Avicenniaceae) Detected by AFLPs and SSRs. *Theoretical and Applied Genetics*, **104**, 388-398. <http://dx.doi.org/10.1007/s001220100724>

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