

Comparison of Positions of QTLs Conferring Resistance to *Xanthomonas campestris* pv. *campestris* in *Brassica oleracea**

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ABSTRACT

Black rot, caused by *Xanthomonas campestris* pv. *campestris* (*Xcc*) is possibly the most important disease of *Brassica oleracea* worldwide. To compare chromosomal positions of *Xcc* resistance loci in *Brassica oleracea* between the present and published studies and to develop marker assisted selection (MAS) to resistance against *Xcc* race 1, we constructed a *B. oleracea* map, including pW, pX and BoCL markers that were closely linked to previously reported *Xcc* resistance QTLs. We also analyzed *Xcc* resistance QTLs by improving our previously reported map derived from the cross of a susceptible double-haploid line (GC P09) with a resistant double-haploid line (Reiho P01). In the nine linkage groups obtained (C1-C9), the major QTL, *XccBo(Reiho)2*, was derived from Reiho with a maximum LOD score (7.7) in C8. The QTL (LOD 4.4) located in C9, *XccBo(GC)1* was derived from the susceptible GC. The other QTL (LOD 4.4), *XccBo(Reiho)1*, was found in C5. Based on common markers, it was possible to compare our finding *Xcc* resistance QTLs with the *B. oleraceaXcc* loci reported by previous authors; *XccBo(Reiho)1* and *XccBo(GC)1* may be identical to the *Xcc* resistance QTLs reported previously or a different member contained in the same resistance gene cluster. Our map includes public SSR markers linked to *Xcc* resistance genes that will promote pyramiding *Xcc* resistance genes in *B. oleracea*. The present study will also contribute to a better understanding of genetic control of *Xcc* resistance.

Keywords: Black Rot; Disease Resistance; QTL; *Xanthomonas campestris* pv. *campestris*

1. Introduction

Black rot, caused by the bacterium *Xanthomonas campestris* pv. *campestris* (Pam.) Dowson (*Xcc*), is the most destructive disease in crucifer crops [1]. *Xcc* enters leaves not only through insect, or mechanically wounded tissue [2] but also through hydathodes at leaf margins and spreads through vascular tissue, clogging vessels and producing V-shaped chlorotic lesions [3]. Such symptoms lead to a systemic infection in susceptible plants so that crop quality and yield substantially decrease. Crop debris and cruciferous weed are potential inoculum sources in field [4]. The pathogen can be retained in seeds via vessels and causes severe incidence in descent

seedlings; consequently, *Xcc* is difficult to prevent by agricultural practices such as seed treatment, crop rotation and use of agrochemicals. Thus, utilization of *Xcc* resistant cultivars is one of the most effective approaches to minimize crop loss from infection of the pathogen.

Nine races of *Xcc* have been identified to date from pathogenicity tests based on the interaction between differential cultivars and races [5-9]. [7] reported that the appearance of race 1 and 4 was predominant worldwide and other races, 2, 3, 5 and 6, were rare. Races 1 and 4 are the most important races in *B. oleracea* crops. Therefore, resistance to both of these two races is a minimum requirement to be of value in controlling black rot [10].

Screening for *Xcc* resistance was performed in *Brassica* species and related species [11-14] and extensive screening using more than 100 genotypes was done by [8]. As a result, resistance to *Xcc* has been identified in

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genotypes of *B. rapa* (A genome), *B. nigra* (B), *B. oleracea* (C), *B. carinata* (BC), *B. juncea* (AB), and *B. napus* (AC). Some genotypes of *B. nigra*, *B. carinata* and *B. juncea* with B genome revealed the highest level of resistance to races 1, 3 and 4, indicating the existence of R1, R3 and R4 resistance genes that were postulated based on the gene-for gene model [8]. In addition, the high level resistance of race 4, conferred by the R4 gene, was found in *B. rapa* and *B. napus* with A genome. In *B. oleracea*, resistance to race 3 and race 5 is common, but resistance to race 1 is very rare. Overall single R genes corresponding to AVR genes (avirulence gene) in each race are considered to confer the high level of qualitative resistance in *Brassica* species, and those R genes are dominant. Inheritance of such race-specific resistance genes was confirmed by the phenotypic segregations in the F2 progenies derived from crossing between susceptible and resistant genotypes; the observed ratio (Resistance:Susceptibility) fits to the 3:1 ratio expected in segregation of a single dominant gene [15].

On the other hand, [11] found that the Japanese cabbage cultivar, Early Fuji, had a high level of resistance to *Xcc*. and he showed that this resistance was controlled by one or two dominant genes. [16] found that resistance of cabbage cultivars BI-16, derived from Early Fuji, was quantitative under oligogenic control; they postulated one major recessive resistance gene, *f*, the expression of which in heterozygous conditions was influenced by one recessive and one dominant modifier genes. [15] also reported that the resistance of BI-16 to race 1 was quantitative and recessive. [17] identified the two *Xcc* resistance QTLs on LG2 and another two QTLs on LG1 and LG9, respectively, in the mapping population derived from the cross of the resistant cabbage and the susceptible broccoli. [18] also identified QTLs on C2, C4, and C5. [19] detected the two significant QTLs controlling resistance to *Xcc* on LG2 and LG9. Those results indicate that resistance to *Xcc* was under oligogenic control. Comparison of the QTLs identified by previous authors is, however, quite difficult because no anchor markers can align the linkage maps constructed by different authors, and furthermore, some of the linkage maps do not follow the international nomenclature established for the C genome of *B. oleracea*.

In our previous paper [19], the total length of the linkage map constructed was 320 cM, which is not long enough to analyze locations of QTLs at a genome-wide level. The objectives of this study, therefore, were 1) to analyze *Xcc* resistance QTLs by using an improved F2 population map of *B. oleracea* plants, and 2) to do a comparative analysis between our mapped QTL positions and the positions of QTLs on the previously published maps by incorporating common markers in our deve-

loped map.

2. Materials and Methods

2.1. Plant Materials

A doubled hybrid (DH) broccoli line (P09) of *B. oleracea* subsp. *Italica* cv. Green Comet (GC) (Takii Seed Co. Ltd., Japan) was crossed as the female parent to a DH line P01 of *B. oleracea* subsp. *capitata* cv. Reiho (Ishii seed company, Japan). The “GC P09” was susceptible to *X. campestris* pv. *campestris* diseases, whereas the Reiho P01 was tolerant. We used the F2 population produced in the study of [19] to construct our linkage map. In summary, seeds of F2 were produced by bud-selfing of a F1 plant and F2 plants selected for QTL analysis were self-pollinated to produce F3 lines. Marathon F1 (*B. oleracea*), Tokyo Cross Hybrid Turnip (*B. rapa*), Seven Top Turnip (*B. rapa*), and Florid Broad Leaf (*B. juncea*) were collected from Twilley Seed Co., Inc., SC, US) and used for identification of the race of *Xcc*. Marathon F1 was used as a susceptible check.

2.2. Inoculation Test and Identification of the Race of *Xcc*

The strain that was used for this study, *X. campestris* pv. *campestris* strain (isolate no. 03-01967), was the same as the report of [19]. Inoculum of the bacterium was cultured in Yeast Dextrose Calcium Carbonate (YDC) agar plate for 48 h at 28°C, and then bacterial cells were scraped from plates and adjusted to a concentration of 10⁸ CFU/ml (0.2OD A600 nm) with 0.85% NaCl solution. V-shaped lesion area (cm²) was measured two weeks after inoculation according to the equation of (lesion width × length) × 1/2.

The seedlings and plants were grown in a greenhouse at the agricultural field of Niigata University. Approximately 50-day old plants were used for the inoculation test. Leaves were inoculated by cutting the mid vein near the leaf margins 1.0 cm in width using a nail cutter that had been dipped in the bacterial suspension [20]. For every inoculation, the nail cutter was dipped into the bacterial suspension. Twelve plants from each F3 line were tested. The three youngest fully expanded leaves were inoculated per plant. The mean DLA of the 12 seedlings per F3 line was used as representative of DLA for each F2 plant.

Identification of race was according to the following criteria; race 0 infected all of four differential cultivars, race 1 infects Marathon F1, Tokyo Cross Hybrid Turnip, and Seven Top Turnip but not Florid Broad Leaf, race 2 infects Marathon F1, Tokyo Cross Hybrid Turnip, and Florid Broad Leaf but not Seven Top Turnip, race 3 infects Marathon F1 and Seven Top Turnip but not

Tokyo Cross Hybrid Turnip and Florid Broad Leaf, race 4 infects only Marathon F1 [5,7].

2.3. DNA Polymorphism and QTL Analysis

The sample set of Genomic DNA of the parents and 94 F2 individuals used in the study of [19] was also used for the linkage construction. DNA of each sample was amplified by the GenomiPhi V2 DNA Amplification kit (GE Healthcare). Ten μ l PCR cocktail containing 10 ng genomic DNA, 0.2 μ M each primer and 1 \times EmeraldAmp Max PCR Master Mix (Takara Bio. Inc., Japan) were used for CAPS (Cleaved Amplified Polymorphic Sequences) and SSR (Simple Sequence Repeat) analyses. Standard three step PCR was performed. Annealing temperature and extension time for PCR were set according to the primer sequence and gene size. The primer sequences were taken from various reports listed in **Table 1**. For CAPS analysis, the amplicons were digested with one of four restriction enzymes (*AluI*, *MspI*, *HinfI* or *MboI*). DNA fragments obtained from restriction enzyme digestion and PCR were separated on 8% - 15% polyacrylamide gel according to fragment size [30]. The gel was subsequently stained with a Gelstar solution (0.1

μ l/10ml) (Takara Bio. Inc., Japan).

Linkage analysis was performed using Ant Map programme, version 1.2 [31]. The QTL detection for *X. campestris* pv. *campestris* resistance was analyzed using a QTL Cartographer software version 2.5 [32] in which composite interval mapping(CIM). CIMs were performed at LOD threshold values which were estimated by means of a permutation test with 1000 permutations with QTL Cartographer.

2.4. Alignment of Different Maps

Previous studies identified four *Xcc* resistance QTLs [17] and self-incompatibility locus [33] in the cabbage BI-16 \times broccoli OSU Cr-7 mapping population, of which linkage map was constructed using WG, TG, and EC RFLP markers. Those markers were renamed as pW and pX according to the NCBI DNA data base. In order to correlate all the linkage groups of the BI-16 \times OSU Cr-7 map to the international nomenclature established for the C genome, we aligned the BI-16 \times OSU Cr-7 map to the consensus map constructed by [29]. After that, common pW and pX markers were used as anchor markers for map position comparisons of *Xcc* resistance QTLs. DNA sequences of the pW/pX RFLP markers were collected from NCBI DNA data base, and then, by using those DNA sequences, we did BLAST search at the *Brassica oleracea* Genomics Project web site, *Bol*base, to obtain coding sequences or genomic sequences corresponding to the RFLP markers. From the identified DNA sequences, we designed primer sets by which the chromosomal regions specifically associated with the pW/pX markers were detected (**Table 2**). For comparison of positions of QTLs identified by [18] and our map, the primer sets were designed based on the sequence of the EST-SNP markers that [18] described. By using the primer set, we amplified the chromosomal region identified by EST-SNP markers and, thereafter, detected the polymorphism by CAPS or PCR-SSCP analysis.

3. Results

For race identification, the lesions on the susceptible differential cultivars and GC P09, enlarged towards the midrib, resulting in typical chlorotic V-shaped lesions (**Figure 1**). The V-shaped lesion was regarded as a triangle shape to calculate disease leaf area (DLA). The lesion on the resistant cultivars and the parent, Reiho P01, was restricted in the portion of leaves that was inoculated by nail cutter with the bacteria suspension after 15 days of inoculation; indicating typically resistant to black rot disease. As a result, among the differential cultivars, only Florida broad leaf (mustard) was found to be resistant to the race (isolate no. 03-01967). This type of reaction

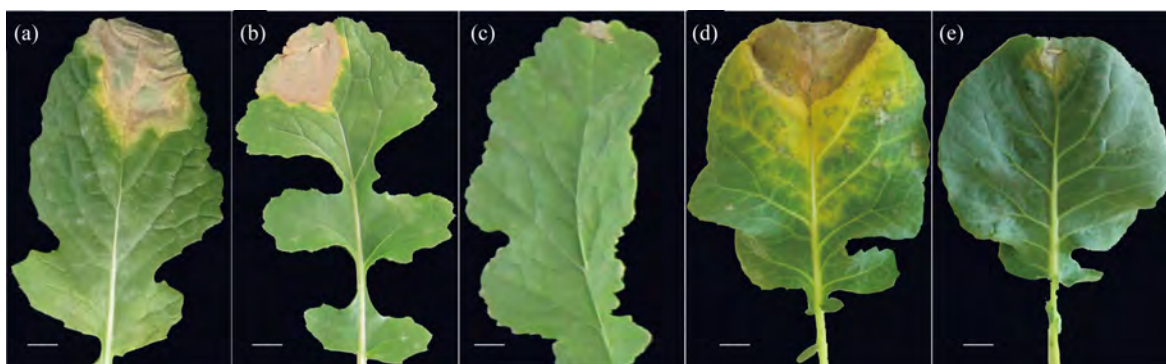
Table 1. List of DNA markers used in this study.

Marker symbols	Type of marker	Origin	Ref.	No. of marker
BnGMS	SSR	<i>B. napus</i>	[21]	1
BoCL	SNP	<i>B. oleracea</i>	[18]	5
BoE	SSR	<i>B. oleracea</i>	[22]	10
BoGMS	SSR	<i>B. oleracea</i>	[23]	43
BrSF	SSR	<i>B. oleracea</i>	[22]	1
BoSF	SSR	<i>B. oleracea</i>	[22]	21 ^a
BRAS	SSR	<i>B. napus</i>	[24,25]	3
BRMS	SSR	<i>B. rapa</i>	[26]	7 ^a
BSA	SSR	<i>B. rapa</i>	[26]	2
CB	SSR	<i>B. napus</i>	[24,25]	34
EMS	SSR	<i>B. oleracea</i>	[22]	3
FITO	SSR	<i>B. oleracea</i>	[27]	1
KBr	SSR	<i>B. rapa</i>	[28]	12
MR	SSR	<i>B. napus</i>	[25]	1
Na	SSR	<i>B. napus</i>	[24,25]	5
Ni	SSR	<i>B. nigra</i>	[24,25]	1
OI	SSR	<i>B. oleracea</i>	[24,25]	2
pW, pX	CAPS	<i>B. napus</i>	[28]	5
	CAPS	<i>B. oleracea</i>	[28]	21 ^b
Total markers				178

^aMarkers mapping more than one position; ^bCAPS markers mapped by using the genotyping data of Doullah *et al.* [19].

Table 2. List of the primer sequences by which the chromosomal regions mapped by pW, PX and BoCL markers were detected.

Name	Forward (5'—3')	Reverse (5'—3')	Chr. identified
BoCL6200s	GGTTGAAAGCAATTGGTGAAC	GGTTCGACACACAAAGAAACCA	C2
BoCL5584	CAAGAGCACAATCTCGGTCTCA	ATGACACGCGTTTACTACTCTGC	C2
pW188	GATGTGATCACCTCTTATCGA	ACAATGCCCCCAACAAAGCG	C3
BoCL5860	AGATGCTACAGCAACAGCTCTC	GAGGAGCTGAGTTGAGAAGCTCA	C5
BoCL1135	TACAAGTACCGCCATAGGTGA	GCATGCTGAAAGATTCTCTGTG	C5
pW114	TTCCCAATGTTGGAGGCAGT	TATATATCGCTCAAGCTCAATC	C5
pW164	CAGCAGCACGATAACGAGGTGCA	CGTGTGATCGTAACGAGCAATTGG	C5
pX117	CGTCCCTTACCTTCTCTCCG	TCCTCCGTAGATAACGGTCCG	C9
pW143	ATGAGCAGAGCACAAAGATCCACCGA	ACAACGGCTTCTCAGAGACCG	C9

**Figure 1.** Black rot disease symptoms produced on differential cultivars and the parent used in QTL analysis. (a) Tokyo Cross Hybrid Turnip; (b) Seven Top Turnip; (c) Florida Broad Leaf; (d) GCP09; (e) Reiho P01. Bar = 1 cm.

between the differential cultivars and the isolates (see Materials and Methods) revealed that the race used in this study was race 1 (Table 3).

In the present study, the inoculation data was cited from [19] who reported that DLA of the F3 lines showed a continuous distribution pattern, with some F3 lines showing lower DLA values than the resistant parent (Figure 2). This time the genotyping of each F2 plant at the *Xcc*-resistance QTL (*XccBo(Reiho)2*) was newly conducted to analyze association between phenotypic and genotypic data of F2 plants (Details given later).

In this study, 94 F2 individuals were used for the linkage construction. A total 181 markers were distributed in 9 linkage groups covering 1099.4 cM, and the average interval between markers was 6.1 cM. The linkage map included 155 SSR and 26 CAPS markers (Table 4). To align our map to the internationally accepted *Brassica* map, we used pW, pX, CB, BRMS, BoGMS and BoSF, markers reported by [22-24,26,29,34].

QTL analysis was performed using the appropriate significance threshold calculated in the permutation test (1000 iterations) and we detected three significant QTLs (Table 5, Figure 3). These results indicated that *Xcc* resistance was controlled by an oligogenic system. Three

QTLs for *Xcc* resistance were detected in C5, C8, and C9. The largest QTL effect (LOD of 7.7) for *Xcc* resistance was detected between the loci EMS1010 and CB10419 on C8 and was closely linked to marker BoGMS0971. This QTL, which explained 34% of the total phenotypic variation, was named *XccBo(Reiho)2*. The QTL located in C9 came from the susceptible broccoli parent (Table 5), and therefore this *Xcc* locus was named *XccBo(GC)1*. Despite high susceptibility of the susceptible parent GC P09 to *Xcc*, this *Xcc* locus accounted for 17.9% of the variation, suggesting that there may be epistatic genes that interact with *XccBo(GC)1* in other regions of the genome. The other minor QTLs found in C5, which came from the resistant parent, accounted for 6.6% of the variation, named *XccBo(Reiho)1*. Genotyping at the BoGMS0971 marker that was closely linked to the major QTL indicated that higher resistance was associated with the homozygous Reiho genotype versus the homozygous GC genotype, with the heterozygotes having varying resistance levels (Figure 2).

For comparison of the positions of *Xcc* resistance QTLs identified by different authors [17,18], we first compared the linkage map (I) of [17] with the linkage map (II) of [33] who mapped self-incompatibility locus.

Table 3. Race identification by using differential cultivars.

Differential cultivars ^a	DLA	Sus/res ^b
Marathon (<i>B. oleracea</i>)	3.1	+
Tokyo Cross Hybrid Turnip (<i>B. rapa</i>)	6.7	+
Seven Top Turnip (<i>B. rapa</i>)	5.8	+
Florida Broad Leaf (<i>B. juncea</i>)	0.02	-

^aDifferential cultivars were chosen from [5] and [7]; ^bSusceptibility (+) /resistance (-).

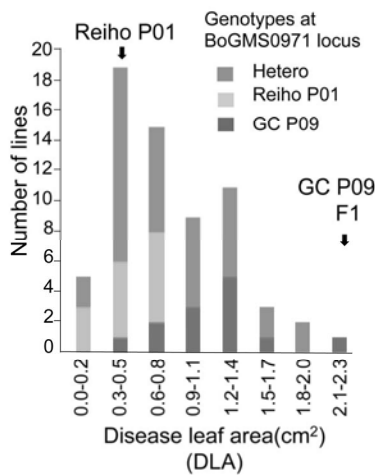


Figure 2. Genotyping data at BoGMS0971 locus in the mapping population that Doullah *et al.* (2010) described previously. Frequency distribution data of mean diseased leaf area (DLA) of black rot disease in the F3 lines. Arrows indicate values obtained for parental (Reiho P01 and GC P09) and F1 plants. The mapping population were genotyped at the BoGMS0971 locus. F2 plants homozygous for the ‘Reiho’ BoGMS0971 locus, homozygous for the ‘GC’ BoGMS0971 locus, or heterozygous at the BoGMS0971 locus are indicated by light gray, gray, or medium gray bars, respectively.

Table 4. Characteristics of *B. oleracea* linkage map.

Linkage Group	Length (cM)	Number of markers		
		SSR	CAPS	Total
1	113.7	15	0	15
2	106.8	16	7	23
3	138.5	12	6	18
4	163.6	22	4	26
5	170.1	19	2	21
6	90.0	17	2	19
7	88.2	19	0	19
8	114.2	19	3	22
9	114.3	16	2	18
Total	1099.4	155	26	181

Although they used the same mapping population (BI-16×OSU Cr-7), the constructed linkage group numbering differed; the LG2 in linkage map (I) corresponded to LG2 and LG9 in linkage map (II), and LG9 in linkage map (I) to the upper portion of LG1 in linkage map (II) (**Table 6**). Next, for assigning positions of *Xcc* resistance QTLs identified by [17] to the consensus map constructed by [29], common pW and pX markers in the two linkage maps were used as anchor markers (**Figure 4**). As a result, LG1 and LG9, that had the major QTLs in BI-16×OSU Cr-7 mapping population, were assigned to the lower portion of C3 and the bottom distal end of C9, respectively. The two minor *Xcc* resistance QTLs on LG2 corresponded to C5 and C6, respectively. The markers, pW164 (WG3C5) and pW114 (TG4D2), that were closely linked to one of the *Xcc* resistance QTLs identified in LG2 by [17] were mapped to the central portion of C5 in our map, where *XccBo(Reiho)1* located. Similarly, the markers, pW143 (WG8A9) and pX117 (EC2D9), that were closely linked to the QTL identified in LG9 were mapped to the distal end of C9 in our map where *XccBo(GC)1* was located. The BoCL6244s marker closely linked to the QTL-3 mapped by [18] was mapped in the central region of C5 in our map. The two *Xcc* resistance QTLs on C3 and C9 were detected by [19] who used the same mapping population used for our study. In the present study, we did not detect significant QTLs on the bottom distal end of C3, although there was a LOD peak (LOD = 2.1) in the same region of C3 where [19] found the *Xcc* resistance QTL.

4. Discussion

Previous studies reported that resistance to *Xcc* occurs with race-specific manner in cruciferous plants including common *Brassica* species and such interaction between *Xcc* and its host was controlled by a gene-for-gene relationship [6,15]. Single dominant genes that are highly resistant against *Xcc* races have been commonly found in *B. nigra*, *B. rapa* and their amphidiploid species, whereas a few sources of race-specific resistance have been identified in *B. oleracea*. On the other hand, non-differential resistance has been found in *B. oleracea* [8]; for instance, [15] reported that F1 plants obtained from the cross of resistant cabbage BI-16 × A12Dhd were susceptible and the subsequent F2 plants showed quantitative resistance to *Xcc* races 1 and 3, indicating that resistance was mainly controlled by one recessive gene (*xca6*) or by linked genes. [16] found that resistance of BI-16 was quantitative under oligogenic control. In the GC P09 × Reiho P01 mapping population, appearance of susceptible phenotype in F1 plants and detection of multiple QTLs controlling resistance to *Xcc* ([19], this study)

Table 5. Summary of QTLs significantly detected for black rot disease against *Xcc* race 1 using F2 populations derived from GC P09 × Reihō P01.

QTL name	Chr.	Closest marker	Position (cM)	LOD ^a	Additive effect ^b	Dominance effect ^b	R ² (%) ^c
<i>XccBo(Reiho)1</i>	C5	BoGMS1330	90.7	4.4	-0.3	0.4	6.6
<i>XccBo(Reiho)2</i>	C8	BoGMS0971	58.0	7.7	-0.9	0.1	34.0
<i>XccBo(GC)1</i>	C9	CB10459	81.0	4.4	0.5	0.0	17.9

^aPeak LOD score of the QTL; ^bAdditive and dominant effect of resistant parent allele in DLA; ^cPercentage of variance explained by quantitative trait loci.

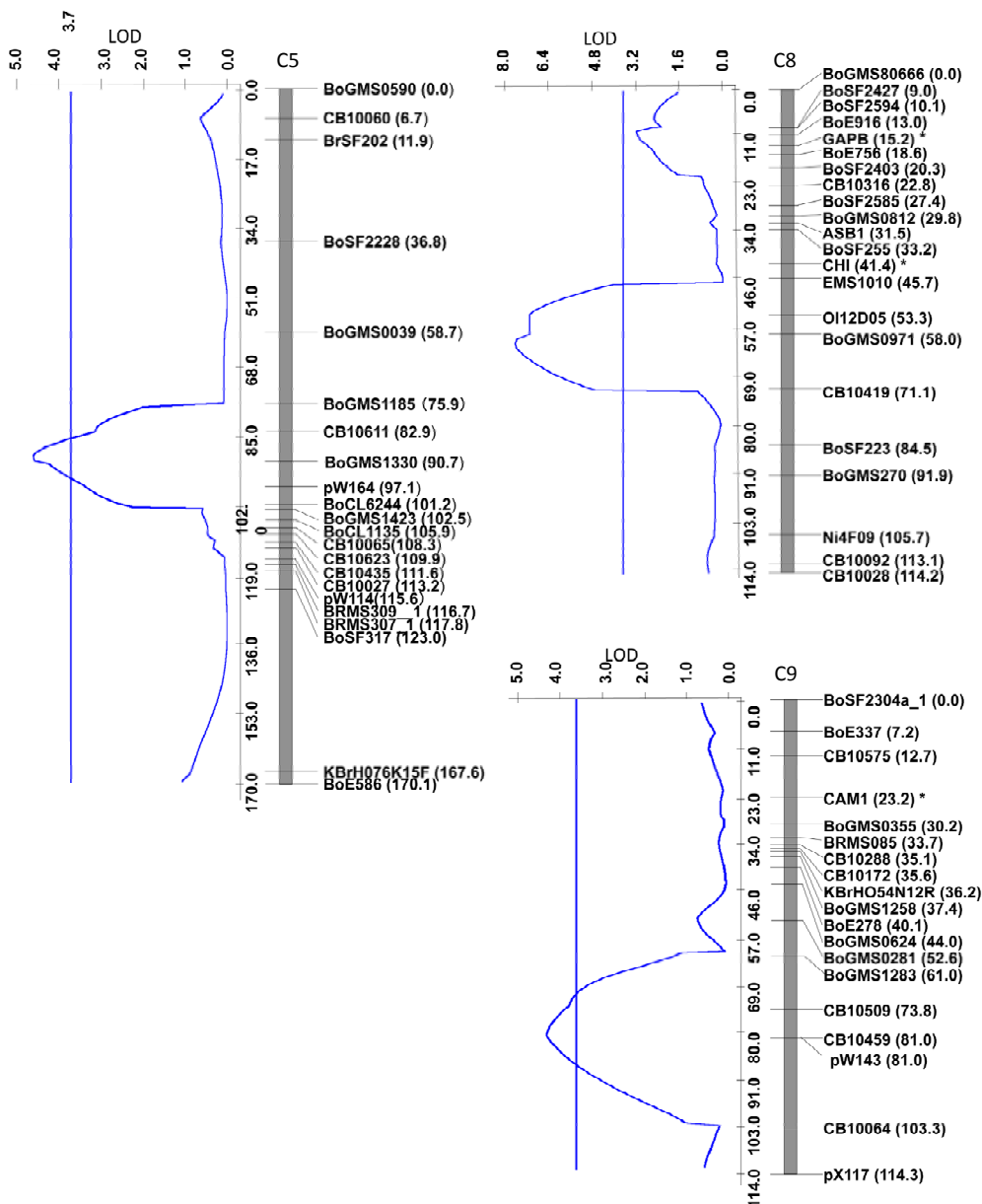


Figure 3. Linkage map developed in the segregating F2 population of broccoli GC (P09) × cabbage Reihō (P01), and LOD profiles for *Xcc* resistance. LOD score profiles and the threshold level (3.6) are shown by blue lines. Linkage groups (O5, O8, O9) that internationally agree with *B. oleracea* reference linkage group nomenclature are indicated at the top of each linkage group. Locus names are indicated on the right side of linkage groups, and map distances in centimorgans are shown in parentheses. CB, BoSF, BoE, BRMS, Ni, Ol, BoGMS, and KBrH markers are cited from various authors (Table 1). The CAPS markers shown by asterisks were mapped by using the data of Doullah *et al.* [19].

Table 6. Alignment of linkage maps for assigning positions of *Xcc*-resistance QTLs identified by Camargo *et al.* (1995) on the consensus map.

Consensus map (Udall <i>et al.</i> [29])	Linkage map (II) (Camargo <i>et al.</i> [33])	Linkage map (I) (Camargo <i>et al.</i> [17])	Marker interval in which <i>Xcc</i> -resistance QTL detected in the linkage map (I)	Abbreviation of the QTLs
N11(C1)	LG8	-		
N12(C2)	LG7	-		
Top of N13 (C3)	LG5	-		
Bottom of N13 (C3)	Bottom of LG1	LG1	WG2G11(pW245)-WG6G5(pW224) ^a WG1E3(pW188)-WG6G5(pW224) ^b	QTL-LG1
N14(C4)	LG4	-		
N15(C5)	LG9	LG2	WG6H1(pW245)-TG4D2(pW114) ^a	QTL-LG2a
N16(C6)	LG2	LG2	EC5E12(pX130)-EC2H2(pX144) ^a	QTL-LG2b
N17(C7)	LG3	-		
N18(C8)	LG6	-		
N19(C9)	Top of LG1	LG9	WG8A9(pW143)-WG4D7 ^a WG8A9(pW143)-EC2D9(pX117) ^b	QTL-LG9

^aThe marker intervals were identified in the greenhouse trial (young plant); ^bin field trial (adult plant).

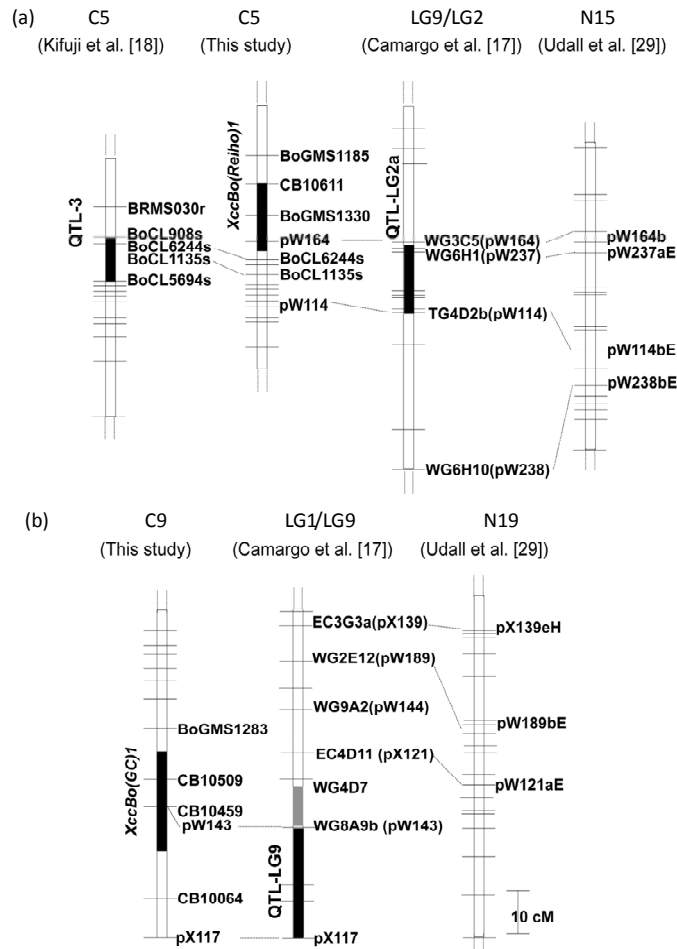


Figure 4. Comparison of chromosomal positions of *Xcc* resistance QTLs on C5 (a) and C9 (b). The filled vertical bars indicate the marker intervals where the *Xcc* resistance QTLs identified by Camargo *et al.* [17], Kifuji *et al.* [18] and this study. The gray and black vertical bars on LG1/LG9 represent QTLs identified by Camargo *et al.* [17] in the greenhouse and the field trials, respectively. The numbers at the top indicate *B. oleracea* reference linkage groups. Consensus map constructed by Udall *et al.* [29] (right) was aligned to identify linkage groups constructed by Camargo *et al.* [17]. Horizontal bars indicate the positions of markers including in each map and the marker names were omitted.

showed that the inheritance of resistance to *Xcc* was recessive and controlled in a quantitative manner.

Taken together, both single (qualitative) and multiple (quantitative) resistance genes must control resistance to *Xcc* in *B. oleracea*.

Our linkage map was constructed from the 94 F₂ plants derived from the cross of GC P09 × Reiho P01 and comprises nine linkage groups, corresponding to the nine chromosomes of *B. oleracea*. The map length of 1099.4 cM was similar to the map length, 1112, 1048, and 1197.9 cM of [22,28,35], respectively, and longer than the map lengths, 891.4, 320.5, 928.7 cM of [18,19,27], respectively. In our map, the average interval between markers was 6.1 cM, indicating that the length and marker distribution of our map were suitable for QTL analysis of *Xcc* resistance. In our study, the QTL analysis that was performed using the appropriate significance threshold successfully detected several significant QTLs, indicating that *Xcc* resistance was controlled by an oligogenic system. We detected one major locus *XccBo(Reiho)2* on C8 that accounted for 34% of the variation. Alleles from Reiho at the *XccBo(Reiho)2* locus act additively and contribute to resistance, as indicated by the negative value of the additive gene action. Genotyping at the BoGMS0971 marker that was closely linked to *XccBo(Reiho)2* showed that the homozygous Reiho genotypes were resistant, whereas the homozygous GC genotypes tended to be susceptible, and the resistance level of the heterozygotes fluctuated (**Figure 2**). This result suggests that the *Xcc* resistance gene at the *XccBo(Reiho)2* locus established stable expression in homozygous plants. We identified two smaller QTLs, *XccBo(Reiho)1* and *XccBo(GC)1*, on C5 and C9, respectively. The resistance allele at *XccBo(GC)1* locus on C9 comes from the susceptible parent. This secret gene effect could explain the fact that some plants exhibited transgressive segregation beyond the level of the resistant parent. In fact, the five most resistant F₃ lines were derived from F₂ plants which were either homozygous for broccoli alleles or heterozygous at this locus on C9. The disease resistance alleles coming from susceptible parents have been reported in QTL analyses of black rot [17] and clubroot [28] in *B. oleracea* and leaf blight in *Zea mays* [36].

[17] identified the four *Xcc* resistance QTLs in the BI-16×OSU Cr-7 map in which two QTLs on LG1 (abbreviation, QTL-LG1) and LG9 (QTL-LG9) were associated with both young and adult plant resistance and the two additional QTLs on LG2 (QTL-LG2a, QTL-LG2b) were associated only with young plant resistance. [18] detected the three *Xcc* resistance QTLs on C2, C4 and C5. Comparative map data with the common pW, pX and BoCL markers revealed that the central portion of C5

harboring *XccBo(Reiho)1* corresponded to that of QTL-LG2a identified by [17] as well as to that of QTL-3 identified by [18]. Furthermore, the bottom distal end of C9 harboring *XccBo(GC)1* corresponded to that of QTL-LG9 identified at the interval pX117 (EC2D9)-pW143 (WG8A9). The assignment of different linkage maps with the common markers suggests that our identified QTL, *XccBo(Reiho)1*, may be equivalent to QTL-LG2a and QTL-3 identified by [17] and [18], respectively, and furthermore, the *XccBo(GC)1* may correlate to QTL-LG9 identified by [17]. The QTL-LG1 in the BI-16×OSU Cr-7 map corresponded to the bottom distal end of C3 based on the location of pW125, pW181, pW245 and pW188 markers (data not shown). We also mapped pW188 on the distal end of C3 in our linkage map where one candidate QTL (LOD = 2.1) was detected. However, it is difficult to conclude whether the QTLs that are linked to the same molecular markers involve just one gene or are family members of clustered *Xcc* resistance genes. Micro-synteny analysis in these regions in *B. oleracea* is needed to identify the relationship between these resistance loci.

Although we identified the *XccBo(Reiho)2* on C8, previous authors did not find any *Xcc* resistance QTLs on C8 [17-19]. The discrepancy in positions of the detected QTLs might be due to differences of races used, inoculation methods, and plant materials. In addition, the magnitude of QTL effects could change in response to different environmental conditions. The upper part of C5, one of middle part of C8 and the distal end of C8 shared conserved regions with *Arabidopsis thaliana* chromosome 1 [37,38]. In addition, it is known that the large conserved regions are duplicated in C5 and C8 [38,39]. This indicates that *XccBo(Reiho)1* on C5 and *XccBo(Reiho)2* on C8 might be homologous loci. The diploid *Brassica* genome was formed by the whole-genome triplication followed by multiple chromosome rearrangements via insertions, deletions, and translocations. Through this process, disease resistance genes were located on various chromosomal regions and furthermore, clustering of disease resistance genes occurred as a result of long-term host-parasite co-evolution [40]. The QTL regions conferring *Xcc* resistance to *B. oleracea* plants might have originated from common chromosomal regions existing in the progenitor diploid species.

Screening genetic resources of *B. oleracea* revealed that resistance to race 3 and race 5 is common, but resistance to race 1 is very rare [8]. Therefore, our finding three QTLs that conferred resistance to race 1 is important for resistant breeding in *B. oleracea*. Pyramiding those QTLs, one novel major QTL on C8 and the other two QTLs that might coincide with previously mapped QTLs, will promote *Xcc* resistance breeding in *B. oleracea*, and the markers closely linked to the QTLs will be

useful in MAS for improving resistance to black rot across environments.

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